

*International Journal of Plant & Soil Science*

*Volume 36, Issue 8, Page 45-56, 2024; Article no.IJPSS.119722 ISSN: 2320-7035*

# **Effect of Transient Waterlogging Stress on Growth, Physiology and Yield of Cowpea (***Vigna unguiculata* **L.) in Semi-Arid Region**

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## *Article Information*

DOI[: https://doi.org/10.9734/ijpss/2024/v36i84834](https://doi.org/10.9734/ijpss/2024/v36i84834)

#### **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/119722>

> *Received: 10/05/2024 Accepted: 12/07/2024 Published: 16/07/2024*

*Original Research Article*

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*Cite as: Senthamil, E., S.S. Angadi, Hanamant M. Halli, S.R. Salakinkop, M.B. Doddamani, and S.S. Gundlur. 2024. "Effect of Transient Waterlogging Stress on Growth, Physiology and Yield of Cowpea (Vigna Unguiculata L.) in Semi-Arid Region". International Journal of Plant & Soil Science 36 (8):45-56. https://doi.org/10.9734/ijpss/2024/v36i84834.*

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# **ABSTRACT**

Cowpea cultivation during rainy season is highly affected by the waterlogging stress due to unpredicted high-intensity rains. The studies on assessment of waterlogging effect on different growth stages of cowpea are necessary for planning mitigation strategies. Hence study was conducted during *kharif* (June to September) 2022 under factorial randomized block design (FRCBD) set up. The first factor consisted of seven waterlogging durations (3 to 15 days), and second factor was three growth stages of cowpea (15 DAE; Days after emergence, 25 DAE and at 50% flowering). The results revealed that regardless of growth stages, growth and yield attributes were drastically decreased with increased duration of waterlogging. The highest plant height (25.07 cm), number of branches plant<sup>-1</sup> (5.33) and leaf area (205.27 cm<sup>2</sup> plant<sup>-1</sup>), and number of pods plant<sup>-1</sup> (4.27), pod length (15.24 cm), number of seeds pod<sup>-1</sup> (14.27), grain (6.27 g plant<sup>-1</sup>) and haulm yield (15.62 g plant<sup>-1</sup>) were recorded with 3 days of waterlogging, whereas lowest values were reported with 15 days of waterlogging. Regarding growth stages, highest growth, and yield attributes were recorded with waterlogging during 50% flowering, followed by 25 DAE and 15 DAE. Moreover, the correlation study indicated that physiological parameters such as leaf protein content  $(r = 0.95)$  and Normalized Difference Vegetation Index NDVI  $(r = 0.97)$  were positively related to grain yield. It was found that, cowpea is sensitive to high-intensity waterlogging (beyond 3–5 days) especially during the early growth stage (15 DAE).

*Keywords: Cowpea; growth stages; protein; proline; waterlogging; yield.*

#### **1. INTRODUCTION**

Cowpea (*Vigna unguiculata* L.) belonging to the family fabaceae acts as a source of livelihood for farmers in the semi-arid regions [1]. Being vegetable meat, cowpea is a rich source of protein, fiber, amino acids, antioxidants, folic acid, phenols and other essential minerals [2]. It is cultivated mainly in countries of semi-arid regions such as Africa, India and Sri Lanka. Globally cowpea is cultivated in an area of around 15 m ha, with production of 9 m t and productivity of  $6 q$  ha<sup>-1</sup> [3]. It is mostly cultivated under rainfed conditions in dry regions as it facilitates shorter life cycle with less water requirement [4]. In India, cowpea is cultivated in an area of 1.6 lakh ha, with the production of 1.03 lakh tonnes and productivity of 8 q ha<sup>-1</sup>. It is a minor pulse crop cultivated mainly in the states of Andhra Pradesh, Karnataka, Tamil Nadu, Telangana, Mizoram, Nagaland and Tripura. The major area (1.01 lakh ha) of cowpea cultivation is under rainfed conditions during *kharif* [5]. Increased summer rainfall saturates the soil before sowing. Waterlogging due to frequent higher intensity rainfall during *kharif* affects the cultivation of cowpea, causing reduction in growth and yield. Waterlogging alters the soil physical and electrochemical properties. It reduces the oxygen diffusion in the soil by 10000 times causing hypoxia and anoxia stress to plants. It also reduces nutrient uptake due to reduced root activity [6].

The associated stress tolerance mechanisms protect the plants from waterlogging effects up to certain duration. The recent study by Basavaraj et al. [7] indicated that waterlogging for 10 days during the early seedling stage reduced the cowpea grain yield by 39.18%. Moreover, Olorunwa et al. [8] indicated that 10 days of waterlogging during vegetative stage reduced the cowpea grain yield by 76%. Whereas according to a previous study by Umaharan et al. [9], waterlogging throughout the crop cycle decreased cowpea grain yield by 53%, while waterlogging at the reproductive stage did not affect yield. Stress tolerance of crops is highly reliant on the genotype, ecotype and adaptive mechanisms. The impact of waterlogging on different growth stages of cowpea remains unclear. The study on the effect of varied durations of waterlogging stress during different growth stages aids in the development of mitigation strategies to obtain higher yield of cowpea. Hence this research was carried out with the objectives to i) assess the effect of varied durations of waterlogging during different growth stages on growth and yield of cowpea, and ii) examine the effect of waterlogging on selected physiological parameters of cowpea and its relation with yield.

#### **2. MATERIALS AND METHODS**

The experiment was conducted during *kharif* 2022 at the ICAR-National Institute of Abiotic

Stress Management, Baramati, Maharashtra, India. The site is located under the scarcity zone of Maharashtra (ACZ-95) with hot and semi-arid climate (AER-6). The experiment was conducted in the pots (capacity: 14 kg soil volume) under factorial randomized block design (FRCBD) with three replications. The durations of waterlogging (D)  $(D_1 - 3, D_2 - 5, D_3 - 7, D_4 - 9, D_5 - 11, D_6 -$ 13 and  $D_7$  – 15 Days) were considered as the first factor and stages of waterlogging (T)  $[T_1 - 15]$ DAE (Days after emergence),  $T_2 - 25$  DAE and  $T_3$  – at 50% flowering stage] were considered as the second factor. Clayey textured soil black was used for the study. The soil was alkaline in reaction (pH: 8.50) with normal electrical conductivity (0.29 dS  $m^{-1}$ ) low in available nitrogen (107.87 kg ha<sup>-1</sup>) and phosphorus (8.16 kg ha-1 ), and medium in available potassium (180.0 kg ha-1 ) and organic carbon content (0.51%). The cowpea variety DC 15 obtained from the University of Agricultural Sciences, Dharwad was sown on 25<sup>th</sup> July 2022. The fertilizer requirement per pot was calculated (Urea: 73.87 mg, DAP: 0.68 g and MOP: 0.25 g) and applied 100% as basal dose on the soil volume basis. Cowpea plants were subjected to waterlogging stress for varied durations at different growth stages by keeping the pots in the constructed concrete tank and maintained the water level of 2.5 cm above soil, whereas, the control pots were maintained with similar management practices except waterlogging stress, where the irrigation was scheduled regularly at 60% of field capacity. The plant growth, physiological and yield parameters were recorded during the course of experiment. The leaf proline content at stress was determined as per the method given by Bates et al. [10] and the leaf protein content was determined as suggested by Singleton et al. [11]. The relationship between plant physiological parameters and yield was studied through Pearson's correlation coefficient analysis. The experimental data were subjected to statistical analysis as outlined by Gomez and Gomez [12]. The critical difference  $(P = 0.05)$  was worked out wherever 'F' test was found significant. Further the mean value of all factors and their interactions were separately subjected to Duncan Multiple Range Test (DMRT) using the corresponding error mean sum of squares and degrees of freedom. Control treatment was analyzed by following Randomized Complete Block Design (RCBD) and presented in the table.

#### **3. RESULTS AND DISCUSSION**

#### **3.1 Effect of Waterlogging on Growth Parameters of Cowpea**

The key growth parameters at 60 days after sowing (DAS) and at harvest were significantly affected by varied durations of waterlogging during the different growth stages (Table 1 and S1).

#### **3.1.1 Plant height (cm)**

Among the durations of waterlogging (D), significantly the highest plant height at 60 DAS (20.86 cm) and harvest (25.07 cm) was recorded with 3 days  $(D_1)$  of waterlogging, whereas the lowest plant height was recorded with 15 days of waterlogging  $(D_7)$ . Similarly, among the stages of waterlogging (T), the plant height (25.92 and 31.88 cm at 60 DAS and harvest respectively) was maximum with waterlogging at 50% flowering and it was followed by waterlogging at 25 DAE (days after emergence) and 15 DAE. Among the interaction effects, plant height at harvest (35.20) was maximum with 3 days of waterlogging from 50% flowering. The minimum plant height (14.19 cm) was recorded with 15 days of waterlogging from 15 DAE. The similar trend was observed at 60 DAS. Waterlogging alters the synthesis, metabolism and transport of endogenous hormones [13]. It results in the inhibition of IAA (auxin), gibberellic acid and cytokinin and increases the accumulation of abscisic acid and ethylene [14]. The reduction in plant height with increased waterlogging duration might be due to the inhibition of the growth promoting hormones [15, 16]. This is attributed to the increase in 1-amino cyclo propane-1 carboxylic acid (ACC), a precursor of ethylene synthesis and signaling molecule induced under hypoxic conditions [17]. It limits the shoot elongation and reduces photosynthesis exclusively at the earlier growth stage [18]. Similarly, Islam et al. [19] reported that five days waterlogging from 15 days after emergence reduced the plant height of mungbean by 28.57% due to the inhibition of growth promoting hormones. In addition to this, nitrogen is necessary for plants vegetative growth. Waterlogging reduces the soil available nitrogen due to denitrification and leaching. The reduced nitrogen availability in the early vegetative growth stage ultimately affects the plant growth. A study by Olorunwa et al. [20] reported that cowpea plant height was reduced due to lesser nutrient uptake attributed to restricted root growth with 10 days of waterlogging stress during vegetative growth stage.

Treatment	<b>Plant Height (cm)</b>		<b>Number of Branches</b> $Plant-1$		Leaf Area $(cm2 plant-1)$	
	<b>60 DAS</b>	At harvest	<b>60 DAS</b>	At harvest	<b>60 DAS</b>	At harvest
Duration of waterlogging (D)						
$D_1 - 3$ Days	$20.86^{\circ}$	25.07a	3.40a	3.53 <sup>a</sup>	615.66a	205.27a
$D_2 - 5$ Days	19.99ab	24.00 <sup>b</sup>	3.06 <sup>b</sup>	3.06 <sup>b</sup>	633.15a	164.66 <sup>ab</sup>
$D_3 - 7$ Days	$19.00^{bc}$	22.76 <sup>c</sup>	2.82c	2.91bc	626.87 <sup>a</sup>	136.56bc
$D_4 - 9$ Days	$18.76^{bc}$	21.71 <sup>d</sup>	2.76 <sup>cd</sup>	2.85 <sup>cd</sup>	565.03 <sup>a</sup>	102.27 <sup>cd</sup>
$D_5 - 11$ Days	$18.59^{bc}$	21.24 <sup>d</sup>	2.62 <sup>cd</sup>	2.70 <sup>de</sup>	555.21 <sup>a</sup>	$91.51c-e$
$D_6 - 13$ Days	$18.72^{bc}$	20.26e	2.57 <sup>d</sup>	2.63 <sup>e</sup>	557.41 <sup>a</sup>	$63.03^{\text{de}}$
$D_7 - 15$ Days	18.38 <sup>c</sup>	19.7 <sup>f</sup>	2.37 <sup>e</sup>	2.41 <sup>†</sup>	556.00 <sup>a</sup>	49.14 <sup>e</sup>
$S.Em. \pm$	0.30	0.232	0.070	0.067	34.81	17.02
Stages of waterlogging (T)						
$T_1 - 15$ DAE*	15.77 <sup>b</sup>	16.56c	2.33c	2.40 <sup>c</sup>	510.86°	129.90 <sup>a</sup>
$T_2 - 25$ DAE	$15.85^{b}$	17.86 <sup>b</sup>	3.31a	3.40 <sup>a</sup>	567.77 <sup>b</sup>	149.68a
50% $\mathsf{T}_3$ $\sim$ $-$ flowering	25.92 <sup>a</sup>	31.88a	2.76 <sup>b</sup>	2.81 <sup>b</sup>	682.51 <sup>a</sup>	68.60 <sup>b</sup>
$S.Em. \pm$	0.52	0.152	0.046	0.044	22.79	11.15
Interaction (DxT)						
$S.Em. \pm$	1.37	0.402	0.122	0.116	60.30	29.49
<b>Control</b>	26.55	35.63	3.66	3.66	760.36	271.33
$S.Em. \pm$	0.79	0.40	0.14	0.13	64.25	43.71
<b>CD at 5%</b>	2.28	1.14	0.41	0.39	183.37	124.75

**Table 1. Effect of waterlogging on growth parameters of cowpea**

*\*DAE, days after emergence; †Means followed by the same letter (s) within the column are not significantly differed (P < 0.05)*

#### **3.1.2 Number of branches plant–1**

Across the growth stages, the number of branches at 60 DAS and harvest (3.40 and 3.53 plant–1 respectively) was maximum with 3 days of waterlogging  $(D_1)$  and minimum (2.37 and 2.41)  $plant^{-1}$  respectively) with 15 days of waterlogging (D7). Among the growth stages, the number of branches was higher with waterlogging at 25 DAE and it was followed by 50% flowering and 15 DAE. Considering the interaction effects, the number of branches at harvest was the highest (4.00 plant–1 ) with 3 days of waterlogging from 25 DAE  $(D_1T_2)$  and the lowest (2.00 plant<sup>-1</sup>) with 15 days of waterlogging from 15 DAE  $(D_7T_1)$ . Similar trend was noted at 60 DAS. The lesser number of branches with waterlogging at early growth stage might be due to the increased energy spent in recovery mechanisms after relieving from waterlogging stress, whereas reduction in number of branches with increased duration of waterlogging can possibly be attributed to the related stress tolerance mechanisms which emphasize only prevention from stress rather than production of newer branches. Our findings are supported by Minchin et al. [21] who reported that waterlogging stressed cowpea plants during

vegetative stage recorded the lowest number of branches  $(9.5 \text{ plant}^{-1})$ . Similarly, 10 days waterlogging stress from 21 days after sowing recorded the lowest number of branches (4.20 plant–1 ) in cowpea [7].

#### **3.1.3 Leaf area (cm<sup>2</sup> plant-1 )**

Considering the waterlogging durations (D), the leaf area at harvest was the highest  $(205.27 \text{ cm}^2)$ plant<sup>-1</sup>) with waterlogging for 3 days ( $D_1$ ) and lowest with 15 days (D7) of waterlogging (49.14  $cm<sup>2</sup>$  plant<sup>-1</sup>). However, the leaf area at 60 DAS was significantly unaffected by the durations of waterlogging. Among the stages of waterlogging (T), the leaf area at 60 DAS was highest (682.51  $cm<sup>2</sup>$  plant<sup>-1</sup>) with waterlogging at 50% flowering. The leaf area at harvest was highest  $(149.68 \text{ cm}^2)$ plant–1 ) with waterlogging at 25 DAE and it was on par with waterlogging at 15 DAE. The leaf area was lowest  $(68.60 \text{ cm}^2 \text{ plant}^{\text{-1}})$  with waterlogging from 50% flowering. Considering the interactions, leaf area at harvest was the highest (218.69 cm<sup>2</sup> plant<sup>-1</sup>) with 3 days of waterlogging from 25 DAE  $(D_1T_2)$ . Whereas, the lowest leaf area (0  $\text{cm}^2$  plant<sup>-1</sup>) was recorded with 15 days of waterlogging from 50% flowering

 $(D_7T_3)$ . The similar trend was observed at 60 DAS. Extended period of waterlogging results in the inhibition of photosynthesis-related enzyme activities, decrease in the ability of leaf chlorophyll synthesis, which causes early senescence, yellowing, and peeling of the leaves and inhibits the growth of newer leaves [22]. This resulted in complete leaf fall at 13 and 15 days of waterlogging from 50% flowering. At harvest, the leaf area was decreased by 52.39% with waterlogging at 15 DAE, 44.83% with waterlogging at 25 DAE and 74.71% with waterlogging at 50% flowering. The reduction in leaf area was in the order of waterlogging at 50% flowering  $> 15$  DAE  $> 25$  DAE. From the interactions, it can be noted that the reduction in leaf area was higher with increased duration of waterlogging from 50% flowering. This was possibly due to the higher translocation of phosynthates to grain under stress conditions which ultimately cause earlier senescence of leaves than waterlogging during other growth stages. Our finding is in line with Ahmed et al. [23], who reported that waterlogging at reproductive stage of mungbean decreased the leaf area by 19.8 to 30.7%.

# **3.2 Effect of Waterlogging on Physiological Parameters of Cowpea**

Waterlogging significantly affected the selected physiological parameters of cowpea at stress (Fig. 1a and b and Table S2).

## **3.2.1 Proline (µmol g–1 )**

Among the waterlogging durations (D), the highest proline content  $(38.85 \text{ \mu mol g}^{-1})$  was recorded with 11 days of waterlogging  $(D_5)$  and remained steady up to 15 days of waterlogging  $(D_7)$ , whereas it was lowest (23.71 µmol g<sup>-1</sup>) with 3 days of waterlogging  $(D_1)$ . Among the stages, the proline content due to varied durations of waterlogging was in the order of 25 DAE > 15 DAE > 50% flowering. Considering the interaction effects, proline content was the highest  $(43.70 \text{ \mu mol g}^{-1})$  with 11 days of waterlogging from 25 DAE (D<sub>5</sub>T<sub>2</sub>). Proline accumulation under waterlogging stress is considered an acclamatory mechanism [24]. It has the functional role of maintaining osmotic adjustment, stabilizing cellular structures and scavenge free radicals during stress [25]. The accumulation of proline with increased duration of waterlogging stress acts as an osmolyte and maintains the plant water status and hydraulic conductivity [26]. Similarly, in our study, the proline content was increased with higher

intensity of waterlogging. The four times higher accumulation of proline due to waterlogging stress was earlier reported in groundnut [27], whereas, 8 days of waterlogging increased the proline content by 101 to 128% in pigeon pea [28].

# **3.2.2 Protein (µg g–1 )**

With regard to waterlogging durations (D), the leaf protein content was the highest  $(31.48 \text{ u})$ <sup>1</sup>) with waterlogging for 3 days  $(D_1)$ , whereas it was lowest  $(20.48 \text{ µg g}^{-1})$  with 15 days of waterlogging (D<sub>7</sub>). However, protein content was significantly unaffected by waterlogging during different growth stages (T). Among the interaction effects, protein content was the highest  $(31.92 \text{ µg g}^{-1})$  with 3 days of waterlogging from 15 DAE  $(D_1T_1)$  and lowest  $(19.7 \mu g g^{-1})$  with 15 days of waterlogging from 50% flowering  $(D_7T_3)$ . Protein content is highly susceptible to varying degrees of stress conditions [29]. Waterlogging stress causes the degradation of plant structural protein due to the dissociation of polyribosomes [30]. The restricted nitrogen uptake due to waterlogging reduces the protein content. Also, the accelerated anoxic metabolism restricts protein synthesis. Therefore, in our study, prolonged duration of waterlogging stress caused a significant decrease in plant protein content. The reduction in leaf protein content with high-intensity waterlogging was earlier reported in wheat [31], pigeon pea [32] and maize [33].

#### **3.2.3 Normalized Difference Vegetation Index (NDVI)**

Among the durations of waterlogging (D), NDVI was highest (0.63) with 3 days  $(D_1)$  and lowest  $(0.45)$  with 15 days  $(D_7)$  of waterlogging. With regard to the stages of waterlogging (T), NDVI values were observed in the trend of 25 DAE  $(0.61)$  > 50% flowering  $(0.52)$  > 15 DAE  $(0.49)$ . Among the interactions, NDVI was highest (0.72) with 3 days of waterlogging from 25 DAE and 50% flowering ( $D_1T_2$  and  $D_1T_3$ ), whereas it was lowest (0.31) with 15 days of waterlogging from 50% flowering (D7T3). NDVI indicates the health and greenness of plant. The increased waterlogging durations reduced the leaf chlorophyll content, subsequently caused yellowing, chlorosis, early senescence of leaves and wilting. This is reflected in the reduced NDVI values with increased waterlogging intensity. The lower NDVI values with waterlogging from 15 DAE were due to lower crop canopy, whereas



#### **Fig. 1. Proline (µmol g–1 ), protein (µg g–1 ) content and NDVI of cowpea at stress as influenced by varied durations (a) and time/stages (b) of waterlogging**

from 50% flowering it was due to higher senescence and leaf fall which resulted in reduced plant canopy. This is in line with the findings of Basavaraj et al. [7] who indicated that waterlogging stressed cowpea plants recorded lower NDVI values (0.38) as compared to nonwaterlogged plants (0.71).

## **3.3 Effect of Waterlogging on Yield Attributes and Yield of Cowpea**

The varied durations of waterlogging during different growth stages exhibited a considerable effect on the yield attributes and yield of cowpea (Table 2 and S3). Among the waterlogging durations (D), higher yield attributes *viz.,* number of pods  $(4.27 \text{ plant}^{-1})$ , pod length  $(15.24 \text{ cm})$  and number of seeds (14.27 pod–1 ) and yield *viz.,* grain  $(6.27 \text{ g plant}^{-1})$  and haulm yield  $(15.62 \text{ g/mol})$ 

plant–1 ) were recorded with 3 days of waterlogging  $(D_1)$  and lower with 15 days of waterlogging  $(D_7)$ . The reduction in grain yield with increased duration of waterlogging can be attributed to higher flower abortion rate and poor pod setting [34, 8]. The stress during 50% flowering produced pods, they elongated and matured within short period. Hence there was less yield loss, though the plants were dried. Across the durations of waterlogging, waterlogging from 50% flowering recorded the higher yield attributes *viz.,* number of pods (3.91 plant–1 ), pod length (15.04 cm) and number of seeds (12.94 pod–1 ) and yield *viz.,* grain (5.14 g plant–1 ) and haulm yield (14.79 plant–1 ). The number of pods  $(2.94 \text{ plant}^{-1})$ , number of seeds  $(12.94 \text{ pod}^{-1})$  and grain yield  $(3.03 \text{ g plant}^{-1})$ were lowest with waterlogging from 15 DAE. On contrary, pod length (13.70 cm).









*\*DAE, days After Emergence; †Means followed by the same letter (s) within the column are not significantly differed (P < 0.05)*





and haulm yield (10.71 g plant–1 ) were lowest with waterlogging from 25 DAE. This indicates that though the plants with waterlogging from 15 DAE produced lengthier pods, it was unable to produce the higher number of seeds pod–1 due to restriction in the source sink relationship which resulted in lesser biomass allocation for seeds leading to chaffy pods. Though the haulm yield was higher with waterlogging from 15 DAE, a lower grain yield was registered. This is linked to the lesser transport of metabolites and nutrients for better seed filling and remobilization of smaller amounts of pre-anthesis resources [35]. This may also be related to the higher energy spent on stress recovery than on reproductive growth [36, 37]. Among the interaction effects (D×T), the higher yield attributes and yield *viz.,* grain (6.89 g plant<sup>-1</sup>) and haulm (19.73 g plant<sup>-1</sup>) yield were recorded with 3 days of waterlogging from 50% flowering  $(D_1T_3)$ . However, the lower yield attributes and yield were recorded with 15

days of waterlogging from 15 DAE  $(D_7T_1)$ . Though the pod length was lower (13.70 cm) with waterlogging from 25 DAE, the number of seeds  $pod^{-1}$  was lesser with waterlogging from 15 DAE (12.94). The restriction in root growth reduces the grain yield of crop [38]. The increased duration of waterlogging especially at early growth stage hinders the root growth and development of roots resulting in reduced nutrient uptake, shoot growth and yield. Furthermore, the production of reactive oxygen species (ROS) due to waterlogging induces damage to the photosystem II activity resulting in reduced photosynthetic activity [39, 40].

#### **3.4 Correlation Studies between Plant Physiological Parameters and Yield**

The correlation studies between plant physiological parameters and yield was significantly affected by waterlogging stress (Fig. 2). The study indicated that the proline content had negative correlation with grain  $(r = -$ 0.94) and haulm  $(r = -0.86)$  yield. This is contrary to the earlier findings that the increased proline content prevents the plants from stress [41,42]. Whereas in this study, the proline content was increased up to 9 days of waterlogging and became steady after that, but the yield was decreased with increased duration of waterlogging. This resulted in the negative association. However, the protein content had positive correlation with grain ( $r =$ 0.95) and haulm ( $r = 0.89$ ) yield. This result is consistent with previous studies that higher protein content is associated with higher nitrogen accumulation [43,44]. Hence the maintenance of higher leaf protein content under waterlogging stress increases the yield during recovery. Similarly, the NDVI had positive correlation with grain ( $r = 0.97$ ) and haulm ( $r = 0.94$ ) yield. The higher NDVI values indicate better plant greenness and health linked with higher yield. This is in line with the findings of Basavaraj et al. [45].

# **4. CONCLUSION**

Cowpea cultivation during rainy season is severely affected by waterlogging due to unpredicted high-intensity rains. Hence, the assessment of cowpea to varied durations of waterlogging at different growth stages is needed. The results suggest that increased durations of waterlogging (3 to 15 days) during different growth stages (15 DAE, 25 DAE and 50% flowering) resulted in the drastic reduction in cowpea growth, and yield. Regardless of waterlogging durations, the lowest growth and yield were recorded with waterlogging at 15 DAE, followed by 25 DAE and 50% flowering. The reduction of key growth and associated physiological parameters reduced the grain and haulm yield. Moreover in this study, the physiological parameters; leaf protein content and NDVI had a positive correlation with yield, while the leaf proline content had negative relationship. From the current investigation, it was found that cowpea is sensitive to highintensity waterlogging (beyond 3–5 days) especially during the early growth stage (15 DAE) over other stages. The findings of this research serve as a guide for the further studies on planning mitigation strategies to reduce the impact of high-intensity waterlogging during the sensitive growth stages of cowpea.

#### **DISCLAIMER (ARTIFICIAL INTELLIGENCE)**

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image generators have been used during writing or editing of manuscripts

# **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## **REFERENCES**

- 1. Halli HM, Angadi SS. Influence of land configuration on rain water use efficiency, yield and economics of cowpea (*Vigna unguiculata* L.) in maize-cowpea sequence cropping under rainfed condition of Northern Transitional Zone. Legume Res - Int J. 2019;42(2):211-215. DOI: 10.18805/LR-3985
- 2. Da Silva AC, da Costa Santos D, Junior DLT, da Silva PB, dos Santos RC, Siviero A. Cowpea: A strategic legume species for food security and health. In: Legume seed nutraceutical research. IntechOpen; 2018.
- 3. Anonymous. Annual report 2022-23. Directorate of Pulses Development, Bhopal, India. 2024;23.
- 4. Halli HM, Angadi SS. Effect of planting methods and deficit irrigation on yield and quality of maize (*Zea mays* L.) stover and their residual effect on cowpea (*Vigna unguiculata* L.) haulm yield on vertisols of semi-arid tropics. Range Manag Agrofor. 2020;41(2):308–315.

DOI: [10.5555/20219973206](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.5555/20219973206)

- 5. Anonymous. Area, Production and Yield– Reports. Directorate of Economics and Statistics, New Delhi, India; 2024. Available[:https://data.desagri.gov.in/websit](https://data.desagri.gov.in/website/crops-apy-report-web) [e/crops-apy-report-web](https://data.desagri.gov.in/website/crops-apy-report-web)
- 6. Armstrong W, 1979, Aeration in higher plants. Adv Bot Res. 1979:7: 225-331.
- 7. Basavaraj PS, Jangid KK, Babar R, Gowdra GVM, Gangurde A, Shinde S, Tripathi K, Patil D, Boraiah KM, Rane J, Harisha CB. Adventitious root formation confers waterlogging tolerance in cowpea (*Vigna unguiculata* (L.) Walp.). Front Sustain Food Sys. 2024:8:1373183.
- 8. Olorunwa OJ, Adhikari B, Brazel S, Bheemanahalli R, Barickman TC and Reddy KR. Waterlogging stress reduces cowpea (*Vigna unguiculata* L.) genotypes growth, seed yield, and quality at different

growth stages: Implications for developing tolerant cultivars under field conditions. Agri Water Manage. 2023;284: 108336.

- 9. Umaharan P, Ariyanayagam RP, Haque SQ. Effect of short-term waterlogging applied at various growth phases on growth, development and yield in *Vigna unguiculata*. The J Agri Sci. 1997; 128(2):189-198.
- 10. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline in water stress studies. *Plant and Soil*, 1973;39:205-208.
- 11. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols, other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods in Enzym.* 1999;299: 152-178.
- 12. Gomez KA, Gomez AA. Statistical Procedure for Agriculture Research, 2<sup>nd</sup> Ed., John Willey and Sons, New York, 680; 1984.
- 13. Chauhan YS, Silim SN, Rao JK, Johansen C. A pot technique to screen pigeonpea cultivars for resistance to waterlogging. J Agron Crop Sci. 1997;178(3):179-83.
- 14. Kyu KL, Malik AI, Colmer TD, Siddique KH, Erskine W. Response of mungbean (cvs. Celera II-AU and Jade-AU) and blackgram (cv. Onyx-AU) to transient waterlogging. Frontiers in Plant Sci. 2021;12:709102.
- 15. Basavaraj PS, Rane J, Boraiah KM, Gangashetty P, Harisha CB. Genetic analysis of tolerance to transient waterlogging stress in pigeonpea (*Cajanus cajan* L. Millspaugh). *Ind. J. Geneti Plant Breed.* 2023;83(03):316-325.
- 16. Lin P, Chen L, Wang W. Mechanisms of mangroves waterlogging resistance. Acta Ecol Sin. 2006;26:586–593. DOI[:https://doi.org/10.3389/fpls.2024.1354](https://doi.org/10.3389/fpls.2024.1354249) [249](https://doi.org/10.3389/fpls.2024.1354249)
- 17. Pan R, Jiang W, Wang Q, Xu L, Shabala S, Zhang WY. Differential response of growth and photosynthesis in diverse cotton genotypes under hypoxia stress. Photosynthetica. 2019;57(3):772–779. DOI: [10.32615/ps.2019.087](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.32615/ps.2019.087)
- 18. Ntukamazina N, Onwonga RN, Sommer R, Mukankusi CM, Mburu J, Rubyogo JC. Effect of excessive and minimal soil moisture stress on agronomic performance of bush and climbing bean (*Phaseolus*

*vulgaris* L.). Cogent Food Agri. 2017; 3(1):1373414. DOI[:https://doi.org/10.1080/23311932.201](https://doi.org/10.1080/23311932.2017.1373414) [7.1373414](https://doi.org/10.1080/23311932.2017.1373414)

- 19. Islam MR, Hasan M, Akter N, Akhtar S. Cytokinin and gibberellic acid alleviate the effect of waterlogging in mungbean (*Vigna radiata* L. wilczek). *Journal Clean WAS*. 2021;5:21-26.
- 20. Olorunwa OJ, Adhikari B, Shi A, Barickman TC. Screening of cowpea (*Vigna unguiculata* (L.) Walp.) genotypes for waterlogging tolerance using morpho-physiological traits at early growth stage. Plant Sci. 2022; 315:111136.

DOI: [10.1016/j.plantsci.2021.111136](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.1016/j.plantsci.2021.111136)

21. Minchin FR, Summerfield RJ, Eaglesham ARJ, Stewart KA. Effects of short-term waterlogging on growth and yield of cowpea (*Vigna unguiculata),* J Agric Sci. 1978;90:355–366.

DOI: [10.1017/S0021859600055465](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.1017/S0021859600055465)

- 22. Wu YS, Yang CY. Physiological responses and expression profile of NADPH oxidase in Rice (*Oryza sativa*) seedlings under different levels of submergence. Rice. 2016;9:2. DOI: [https://doi.org/10.1186/s12284-016-](https://doi.org/10.1186/s12284-016-0074-9)
- [0074-9.](https://doi.org/10.1186/s12284-016-0074-9) 23. Ahmed S, Nawata E, Sakuratani, T. Effects of waterlogging at vegetative and<br>reproductive growth stages on reproductive growth stages on photosynthesis, leaf water potential and yield in mungbean. Plant Production Sci. 2002;5(2): 117-123.
- 24. Barnett NM, Naylor AW. Amino acid and protein metabolism in Bermuda grass during water stress. Plant Phy. 1966;41:1222–1230.
- 25. Delauney AJ, Verma DPS. Proline biosynthesis and osmoregulation in plants. Plant J. 1993;4:215–223.
- 26. Barickman TC, Simpson CR, Sams CE. Waterlogging causes early modification in the physiological performance, carotenoids, chlorophylls, proline, and soluble sugars of cucumber plants. Plants. 2019;8(6):160.
- 27. Vurayai R, Emongor V, Moseki B Physiological responses of bambar groundnut (*Vigna subterranea* L. Verdc) to short periods of water stress during different developmental stages. Asian J Agri Sci. 2011;3:37–43.
- 28. Duhan S, Kumari A, Bala S, Sharma N, Sheokand S. Effects of waterlogging,

salinity and their combination on stress indices and yield attributes in pigeonpea (*Cajanus cajan* L. Millsp.) genotypes. Ind J Plant Phy. 2018;23:65-76.

- 29. Kattimani, KN, Patil BN, Hanchinal RR, Kulkarni VN. Effects of irrigation on yield, protein content and seedling vigour in wheat . J Maharash Agri Univ. 1996;21(2):295–296.
- 30. Kennedy RA, Rumpho ME, Fox TC. Anaerobic metabolism in plants. Plant Phy. 1992;100:1–6.
- 31. Olgun M, Kumlay MA, Adiguzel CM, Caglar A. The effect of waterlogging in wheat (T. *aestivum* L.). Acta Agri Scandi Sec B–Soil and Plant Sci. 2008;58(3):193- 198.
- 32. Bansal R, Srivastava JP. Effect of waterlogging on photosynthetic and biochemical parameters in pigeonpea. Rus J Plant Phy. 2015;62:322-327.
- 33. Tian L, Bi W, Liu X, Sun L, Li J. Effects of waterlogging stress on the physiological response and grain-filling characteristics of spring maize (*Zea mays* L.) under field conditions. Acta Physiologiae Planta*.*  2019;41:1-14.
- 34. Timsina J, Garrity DP, Pandey RK. Plant water relations and growth of cowpea cultivars subjected to varying degrees of waterlogging. Field Crops Res. 1994;39(1):49-57.
- 35. Li C, Jiang D, Wollenweber B, Li Y, Dai T, Cao W. Waterlogging pretreatment during vegetative growth improves tolerance to waterlogging after anthesis in wheat. Plant Sci. 2011;180(5):672–678. DOI:https://doi.org/10.1016/j.plantsci.2011. 01.009.
- 36. Arduini I, Baldanzi M, Pampana S. Reduced growth and nitrogen uptake during waterlogging at tillering permanently affect yield components in late sown oats. Frontiers in Plant Sci. 2019;12(10): 457867.
- 37. Zhang Y, Liu G, Dong H, Li C. Waterlogging stress in cotton: Damage, adaptability, alleviation strategies, and mechanisms. The Crop J. 2021;9(2): 257–270.

DOI:https://doi.org/10.1016/j.cj.2020.08.00 5

- 38. Halli HM, Angadi SS, Kumar A, Govindasamy P, Madar R, El-Ansary DO et al. Influence of planting and irrigation levels as physical methods on maize root morphological traits, grain yield and water productivity in semi-arid region. Agron. 2021;11(2):294.
- DOI: [10.3390/agronomy11020294](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.3390/agronomy11020294) 39. Nishiyama Y, Allakhverdiev SI, Murata N. A new paradigm for the action of reactive oxygen species in the photoinhibition of photosystem II. BBA-Bioenerg. 2006;1757: 742–749.

DOI: [10.1016/j.bbabio.2006.05.013](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.1016/j.bbabio.2006.05.013)

40. Ashraf MA. Waterlogging stress in plants: a review. Afr J Agric Res. 2012;7:1976– 1981.

DOI: [10.5897/AJAR2016.12050](file:///C:/Users/HM%20Halli/OneDrive/Desktop/10.5897/AJAR2016.12050)

- 41. Migdadi HM, El-Harty EH, Salamh A, Khan MA. Yield and proline content of faba bean genotypes under water stress treatments. JAPS: J Animal & Plant Sci. 2016;26(6).
- 42. Arteaga S, Yabor L, Díez MJ, Prohens J, Boscaiu M, Vicente O. The use of proline in screening for tolerance to drought and salinity in common bean (*Phaseolus vulgaris* L.) genotypes. Agron. 2020;10(6): 817.
- 43. Ravelombola WS, Shi A, Weng Y, Motes D, Chen P, Srivastava V et al. Evaluation of total seed protein content in eleven Arkansas cowpea (*Vigna unguiculata* (L.) Walp.) lines. Am J Plant Sci. 2016;7:2288– 2296.

Available:https://doi.org/ 10.4236/ajps.2016.715201.

- 44. Weng Y, Qin J, Eaton S, Yang Y, Ravelombola WS, Shi A. Evaluation of seed protein content in USDA cowpea germplasm. HortScience. 2019;54:814– 817. Available[:https://doi.org/10.21273/HORTS](https://doi.org/10.21273/HORTSCI13929-19) [CI13929-19.](https://doi.org/10.21273/HORTSCI13929-19)
- 45. Basavaraj PS, Jangid KK, Babar R, Rane J, Boraiah KM, Harisha CB et al. Non-invasive measurements to identify mungbean genotypes for waterlogging tolerance. Peer J. 2024;12: e16872.

# **SUPPLEMENTARY MATERIALS**



# Table S1. Effect of waterlogging on growth parameters of cowpea

\*DAE, Days After Emergence; † Means followed by the same letter (s) within the column are not significantly<br>differed (P < 05)





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\*DAE, Days After Emergence; † Means followed by the same letter (s) within the column are not significantly differed  $(P < .05)$ 

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