

American Chemical Science Journal 4(4): 516-536, 2014



SCIENCEDOMAIN international www.sciencedomain.org

## Synthesis, Antiviral and Kinesin Eg5 Activities of New Indomethacin Analogues

Najim A. Al-Masoudi<sup>1\*</sup> and Dawood S. Ali<sup>1</sup>

<sup>1</sup>Department of Chemistry, College of Science, University of Basrah, Basrah, Iraq.

## Authors' contributions

This work was carried out in collaboration between both authors. Author NAI-M designed the study, wrote the protocol, managed the analyses of the study, managed the literature searches and wrote the first draft of the manuscript. Author DSA synthesized the new compounds. Both authors read and approved the final manuscript.

**Original Research Article** 

Received 26<sup>th</sup> December 2013 Accepted 20<sup>th</sup> February 2014 Published 3<sup>rd</sup> March 2014

## ABSTRACT

**Aim:** Synthesis, characterization, anti-HIV, anti-HCV and kinesin activities of new indomethacin analogues have been carried out.

**Methodology:** Arylated derivatives of indomethacin via the Suzuki-Miyaura crosscoupling reaction using palladium acetate/triphenylphophineor palladium based *N*heterocyclic carbene (Pd-NHC) complexe as catalysts were synthesized and characterized by the <sup>1</sup>H and <sup>13</sup>C and 2D NMR study. Analogously, indomethacin analogues bearing thioureido and amide moieties of various L-amino acid esters were prepared *via* Kabbani and coupling reactions, respectively.

**Results:** All the new analogues were evaluated *In vitro* for their antiviral activity against the replication of HIV-1 and HIV-2 in MT-4 cells using MTT assay. Compounds 28, 31 and 32 were evaluated *In vitro* for their inhibitory activity against hepatitis virus C (HCV) in the Huh 5-2 replicon system (type 1b, Con1 strain). Additionally, some analogues were screened for their inhibitory activity against the ATPase enzyme and the motor-protein Kinesin Eg5.In conclusion, Compounds 31 and 39 showed anti-HIV activity with IC<sub>50</sub> values of >1.81 and >  $3.21\mu$ M (CC<sub>50</sub> of 3.31 and  $28.89\mu$ M), resulting in selectivity indexs (SI) of 6 and 9, respectively.

**Conclusion:** Compounds 31 and 39 displayed better anti-HIV activity than the other derivatives (SI = 6 and 9, respectively). Compound 27 showed ATPase inhibition value of 48% at 100  $\mu$ M concentration.

<sup>\*</sup>Corresponding author: Email: najim.al-masoudi@gmx.de;

Keywords: Antiviral activity; anti-kinesin Eg5 inhibitors; indomethacin; suzuki-miyaura crosscoupling reaction; thioureido derivatives.

## **1. INTRODUCTION**

Non-steroidal anti-inflammatory drugs (NSAIDs) are considered to be the most important therapeutic agents for the treatment of pain and inflammation related to a large variety of pathologies like rheumatoid arthritis and osteoarthritis. Such drugs are indomethacin, 1[1,2] etodolac [3] and tenidap [4]. Indomethacin, as one of NSAIDs, blocks the enzymes that make prostaglandins (cyclooxygenase (COX-1 and 2) and thereby reduces the levels of prostaglandin (PG), leukotriene and thromboxane precursors [5-8]. As a result, fever, pain and inflammatory rheumatoid diseases are reduced. Marnett et al. [9-12] reported that substitution of the 2'-methyl group of indomethacin with trifluoromethyl produces CF<sub>3</sub>- indomethacin, a tight-binding inhibitor with kinetic properties, showed unexpected COX-2 selectivity in comparison to indomethacin itself (IC<sub>50</sub> mCOX-2 = 267 nM, IC<sub>50</sub> COX-1 > 100  $\mu$ M). Furthermore, COX inhibitors, indomethacin and its structural analogue sulindac exhibit cell growth inhibition and apoptosis inducing activities in various cancer cell lines *via* COX independent mechanisms. [13] On the other hand, it has been found that removal of the methyl group from 1 would lead drastically in reduction of the inhibitory potency against both COX enzymes [14].

Recently, Kalgutkar et al. [15,16] have reported that derivatization of indomethacin carboxylic acid group to ester or amide analogues produces compounds would lead to highly selective and potent COX-2 inhibitors, while other analogues having N-difluoromethyl-1,2dihydropyrid-2-one moiety [17] reported as cyclooxygenase and lipoxygenase inhibitors. Zorc et al. [18] have examined some NSAID hydroxamic acids against malignant tumor cell lines and normal human fibroblasts and found indomethacin analogue 2 exhibited a moderate inhibition against breast carcinoma, murine leukemia and human T lymphocytes (mean IC<sub>50</sub> = 4.2-28  $\mu$ M). In addition, alkyl-tin carboxylate derived from indomethacin exhibited remarkable In vitro cytotoxicity [19], while some new indomethacin analogues have been reported as potent antitumor agents [20,21], with remarkable cytotoxic activity(e.g. 3) [22]. Unexpectedly, Santoro et al. [23] have found that indomethacin has a potent antiviral activity against the coronaviruses SARS-CoV. In 2005, Richardson et al. [24]have reported that indomethacin is a potential therapeutic target in respiratory syncytial virus (RSV) therapy, meanwhile Mukherjee and Simpson [25] have indicated the effect of indomethacin In vitro on the vesicular stomatitis virus. Furthermore, Bourinbaiar and Lee-Huang [26] have studied the anti-HIV activity of anti-inflammatory drugs, dexamethasone and indomethacin, by MAP30, the antiviral agent from bitter melon.

In continuation of our ongoing work on the synthesis of new anti-HIV agents and our recent antiviral data on new indomethacin derivatives [27], we report here the synthesis of new indomethacin analogues with evaluation of their anti-HIV, anti-HCV and anti-kinesin Eg5 activity.

American Chemical Science Journal, 4(4): 516-536, 2014



## 2. EXPERIMENTAL

## 2.1 Materials and Methods

Melting points are uncorrected and were measured on a Büchi melting point apparatus B-545 (BÜCHI Labortechnik AG, Switzerland). NMR data were obtained 400 and 600 MHz (<sup>1</sup>H) and 150.91 MHz (<sup>13</sup>C) spectrometers (Avance III, Bruker, Germany) with TMS as internal standard and on  $\delta$ scale in ppm. All nmr spectra were measured in dimethyl sulfoxide d<sub>6</sub>. Heteronuclear assignments were verified by <sup>1</sup>H-<sup>13</sup>C COSY, or HMQC experiments. Microanalytical data were obtained with a Vario, Elemental apparatus (Shimadzu, Japan). Mass spectra were recorded on 70 eV EI and FAB MAT 8200 spectrometers (Finnigana MAT, USA). Silica gel (0.040-0.063 mm) used for column chromatography and analytical silica gel TLC plates 60 F254 were purchased from Merck.

## 2.2 Synthesis of the Compounds

2-(1-(4-Chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetyl chloride (4) This compound was prepared according to the reported procedure [15]:

## 2.2.1 General procedure of preparation of indomethacin having triaryl groups via Suzuki-Miyaura cross-coupling reaction [17-27]

*Method A*. A mixture of indomethacin 1 (100 mg, 0.28 mmol) and arylboronic acids 5-16 (0.29 mmol) in *n*-propanol (15 mL) was stirred for 15 min, and to this mixture was added  $Pd(OAc)_2$  (19 mg, 0.085 mmol), triphenylphosphene (66mg, 0.25 mmol) and 2m aq. solution of Na<sub>2</sub>CO<sub>3</sub> (3.5 mL). The reaction mixture was refluxed for 4-6 h and completion of reaction was monitored by TLC. After cooling, water was added (7 mL), followed by stirring for 5 min and the mixture was partitioned with ethyl acetate (3×10 mL). The combined organic extracts were dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and evaporated to dryness. The crude product was purified on a column of SiO<sub>2</sub> (5 g) using: hexane-EtOAc = 3:2 or, in gradient, MeOH (0-10%) and CHCl<sub>3</sub> as eluents to give the desired product.

*Method B.* A mixture of indomethacin 1 (358 mg, 1.0 mmol), arylboronic acids 5-7 (1.2 mmol), Pd-NHC catalyst (1 mol %, 0.0096 g),  $K_2CO_3$  (276 mg, 2.0 mmol), and a mixture of

acetone/water (1:1) (10 mL) was stirred at 40 °C for 3 h The reaction mixture was then diluted with water (20 mL) and extracted three times with dichloromethane (3×10 mL). The combined organic layer was washed with brine (20 mL) and dried (Na<sub>2</sub>SO<sub>4</sub>), filtered and the filtrate was evaporated to dryness. The crude products was purified on a SiO<sub>2</sub> as in method A to give 17-19. The physical data were identical for those compounds prepared previously in method A.

# 2-(1-(2'-Fluoro-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (17):

From 2-fluorophenylboronic acid 5 (39 mg). Yield: 123 mg (88%), as a yellow powder, mp 198-200 °C.<sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$  10.42 (s, 1H, CO<sub>2</sub>H), 7.88 (dd, 2H, *J* = 7.0, 1.8 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.74 (m, 1H, 6''-H<sub>arom</sub>), 7.64-7.49 (m, 3H, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub> + 4''-H<sub>arom</sub>), 4.32 (m, 2H, 3''-H<sub>arom</sub> + 5''-H<sub>arom</sub>), 7.05 (d, 1H, *J* = 8.1 Hz, 7-H<sub>indole</sub>), 6.96 (d, 1H, *J* = 2.1 Hz, 4-H<sub>indole</sub>), 6.54 (dd, 1H, *J* = 8.1, 2.4 Hz, 6-H<sub>indole</sub>), 3.71 (s, 3H, OMe), 3.48 (s, 2H, CH<sub>2</sub>), 2.28 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  176.0 (CO<sub>2</sub>H), 168.9 (C=O), 157.5 (C-OMe + C-F), 142.6 (C4'<sub>arom</sub>), 138.9 (C3a<sub>indole</sub>), 133.5 (C2<sub>indol</sub>), 132.4, 131.4, 130.8, 129.5, 128.6, 127.0 (C<sub>arom</sub>), 124.9 (d, *J*<sub>C,F</sub> = 120 Hz, C1''<sub>arom</sub>), 117.4 (d, *J*<sub>C,F</sub> = 240 Hz, (C3''<sub>arom</sub>), 110.2 (C7<sub>indol</sub>), 108.7 (C3<sub>indol</sub>), 108.3 (C6<sub>indol</sub>), 100.8 (C4<sub>indol</sub>), 55.3 (OMe), 30.7 (CH<sub>2</sub>), 11.6 (Me). Anal. calcd for C<sub>25</sub>H<sub>20</sub>FNO<sub>4</sub> (417.43): C, 71.93; H, 4.83; N, 3.36. Found: C, 71.71; H, 4.73; N, 3.09.

# 2-(1-(3'-Fluoro-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (18):

From 3-fluorophenylboronic acid 6 (39 mg). Yield: 96 mg (82%), as a yellow powder, mp 200-202 °C.<sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$  10.41 (s, 1H, CO<sub>2</sub>H), 7.89 (dd, 2H, *J* = 6.9, 1.9 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.66-7.55 (m, 4H, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub> + 2"-H<sub>arom</sub> + 4"-H<sub>arom</sub>)7.32 (dd, 2H, *J* = 8.3, 2.3 Hz, 5"-H + 6"-H<sub>arom</sub>), 7.06 (d, 1H, *J* = 8.6 Hz, 7-H<sub>indole</sub>), 6.96 (d, 1H, *J* = 2.4 Hz, 4-H<sub>indole</sub>), 6.55 (dd, 1H, *J* = 8.6, 2.4 Hz, 6-H<sub>indole</sub>), 3.71 (s, 3H, OMe); 3.48 (s, 2H, CH<sub>2</sub>), 2.28 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  176.0 (CO<sub>2</sub>H), 168.8 (C=O), 161.8 (d, *J*<sub>C,F</sub>= 257 Hz, C-F), 161.2 (C-OMe), 146.5 (C-4'arom + C1"arom), 139.0 (C3a<sub>indol</sub>), 133.4 (C2<sub>indol</sub>), 132.1, 132.0, 131.4, 130.8, 130.1, 129.4, 128.7, 126.9 (C<sub>arom</sub>), 123.2 (d, *J*<sub>C2",F</sub> = 120 Hz, C2"arom), 116.4 (d, *J*<sub>C4",F</sub>= 120 Hz, C4"arom), 114.8 (C3<sub>indol</sub>),110.2 (C7<sub>indol</sub>), 108.6 (C6<sub>indol</sub>), 100.8 (C4<sub>indol</sub>), 55.2 (OMe), 30.8 (CH<sub>2</sub>), 11.1 (Me). Anal. calcd for C<sub>25</sub>H<sub>20</sub>FNO<sub>4</sub> (417.43): C, 71.93; H, 4.83; N 3.36. Found: C, 71.73; H, 4.69; N, 3.18.

# 2-(1-(4'-Fluoro-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (19):

From 4-fluorophenylboronic acid 7 (39 mg). Yield: 120 mg (86%), as a yellow product, mp250-252°C. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$  10.45 (s, 1H, CO<sub>2</sub>H), 7.90 (dd, *J* = 8.5, 2.2 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.66-7.56 (m, 2H, 2"-H<sub>arom</sub> + 6"-H<sub>arom</sub>), 7.34-7.31 (m, 4H, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub> + 3"-H<sub>arom</sub> + 5"-H<sub>arom</sub>), 7.06 (d, 1H, *J* = 8.4 Hz, 7-H<sub>indol</sub>), 7.05 (d, 1H, *J* = 2.2 Hz, 4-H<sub>indol</sub>), 6.59 (dd, 1H, *J* = 8.4 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.25 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  175.9 (CO<sub>2</sub>H), 168.9 (C=O), 161.5 (d, *J*<sub>C4",F</sub>= 252 Hz, C4"<sub>arom</sub>-F), 160.8 (*C*-OMe), 145.8 (C4'<sub>arom</sub>), 138.9 (C3a<sub>indol</sub>), 133.4 (C1"<sub>arom-F</sub>), 132.4 (C2<sub>indol</sub>), 131.4, 130.8, 129.4, 128.6, 127.0 (C<sub>arom</sub>), 118.1 (2d, *J* = 121 Hz, C3"<sub>arom</sub> + C5"<sub>arom</sub>), 116.0 (C3<sub>indol</sub>), 114.1 (C7<sub>indol</sub>), 110.2 (C6<sub>indol</sub>), 100.0 (C4<sub>indol</sub>), 55.2 (OMe), 33.8 (CH<sub>2</sub>), 11.6 (Me). Anal. calcd for C<sub>25</sub>H<sub>20</sub>FNO<sub>4</sub> (417.43): C, 71.93; H, 4.83; N, 3.36. Found: C, 71.69; H, 4.77; N, 3.12.

# 2-(1-(3',4'-Difluoro-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (20):

From 3,4-difluorophenylboronic acid 8 (44 mg). Yield: 112 mg (78%), as a yellow powder, mp 190-192°C. <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.42 (s, 1H, CO<sub>2</sub>H), 7.89 (dd, 2H, *J* = 8.6, 2.2 Hz, 3'-

Harom + 6'-Harom ), 7.64-7.53 (m, 1H, 6"-Harom-F), 7.32 (d, J = 8.6 Hz, 2H,2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.61 (m, 1H, 6"-H<sub>arom</sub>-F), 7.55 (dd, 2H, J = 8.6, 2.2 Hz, 2'-Harom + 5'-Harom), 7.31 (m, 1H, 5"-H<sub>arom</sub>), 7.06 (d, 1H, J = 8.0 Hz, 7-Hindol), 6.94 (d, 1H, J = 2.1 Hz, 4-Hindol), 6.66 (m, 1H, 6"-H<sub>arom</sub>), 6.54 (dd, 1H, J = 8.0, 2.0 Hz 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.47 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  176.1 (CO<sub>2</sub>H), 168.9 (C=O), 154.1 (C-OMe), 152.4 (C4'<sub>arom</sub>), 148.4 (d,  $J_{C3",F} = 256$  Hz, C3"<sub>arom</sub>), 140.8 (d,  $J_{C4",F} = 255$  Hz, C4"<sub>arom</sub>), 138.8 (C3aindol), 133.4 (C2indol), 132.3, 131.4, 130.1, 129.4, 128.6 (C<sub>arom</sub>), 123.1 (d,  $J_{C2",F} = 122$  Hz, C2"<sub>arom</sub>), 121.5 (d,  $J_{C5",F} = 120$  Hz, C5"<sub>arom</sub>), 116.0 (C3indol), 115.4 (C7<sub>indol</sub>), 110.2 (C6<sub>indol</sub>), 100.9 (C4indol), 30.8 (CH<sub>2</sub>), 55.2 (OMe). Anal. calcd for C<sub>25</sub>H<sub>19</sub>F<sub>2</sub>NO<sub>4</sub> (435.42): C, 68.96; H, 4.40; N, 3.22. Found: C, 68.69; H, 4.32; N, 3.01.

### 2-(1-(3',4'-Dimethoxy-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3yl)aceticacid (21):

From 3,4-dimethoxyphenylboronic acid 9 (51 mg). Yield: 138 mg (91%), as a brown powder, mp 210-212°C.<sup>1</sup>H NMR DMSO- $d_6$ ):  $\delta$  10.42 (s, 1H, CO<sub>2</sub>H), 7.88 (dd, 2H, J = 8.5, 2.3 Hz, 2'-Harom + 6'-Harom ), 7.62 (m, 1H, 2"-H<sub>arom</sub>), 7.32 (dd, 2H, J = 8.5, 2.3 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.09 (d, 1H, J = 8.2 Hz, 7-H<sub>indol</sub>), 7.04 (d, 1H, J = 2.1 Hz, 4-H<sub>indol</sub>), 6.94 (d, 1H, J = 8.0 Hz, 3"-H<sub>arom</sub>), 6.55 (br s., 1H, 6"-H<sub>arom</sub>), 6.53 (dd, 1H, J = 8.2, 2.1 Hz, 6-H<sub>indol</sub>), 3.83, 3.77 (2xs, 6H, OMe), 3.42 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  175.9 (CO<sub>2</sub>H), 168.8 (C=O), 161.6 (C5-OMe), 152.4 (C5"-OMe), 148.9 (C4"-OMe), 147.9 (C4'<sub>arom</sub>), 139.0 (C1"<sub>arom</sub>), 133.3 (C(3a)<sub>indol</sub>), 132.2 (C(2)<sub>indol</sub>), 130.7, 130.1, 127.0 C<sub>arom</sub>), 120.6 (C2"<sub>arom</sub>), 114.3 (C3<sub>indol</sub> + C7<sub>indol</sub>), 112.1 (C6"<sub>arom</sub>), 110.1 (C6<sub>indol</sub>), 108.5 (C3"<sub>arom</sub>), 100.8 (C(4)<sub>indol</sub>), 55.5, 55.2 (3xOMe), 30.8 (CH<sub>2</sub>), 11.6 (Me). Anal. calcd for C<sub>27</sub>H<sub>25</sub>NO<sub>6</sub> (459.17): C, 70.58; H, 5.48; N, 3.05I. Found: C, 70.32; H, 5.40; N, 2.85.

## 2-(5-Methoxy-2-mehyl-1-(2'-(methylthio)-[1,1'-biphenyl]4-carbonyl)-indol-3yl)aceticacid (22):

From 2-methylthiophenylboronic acid 10 (47 mg). Yield: 106 mg (72%), as a colourless powder, mp 238-240 °C.<sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.42 (s, 1H, CO<sub>2</sub>H), 8.11 (dd, 2H, *J* = 8.0, 2.1 Hz, 2'-Harom + 6'-Harom), 7.88 (dd, 2H, *J* = 8.0, 2.1 Hz, 3'-H<sub>arom</sub> + 5'-Harom), 7.58 (m, 2H, 3"-H<sub>arom</sub> + 6"-Harom), 7.31 (dd, 2H, 4"-H<sub>arom</sub> + 5"-Harom), 7.05 (d, 1H, *J* = 8.4 Hz, 7-Hindol), 6.95 (d, 1H, *J* = 2.2 Hz, 4-Hindol), 6.53 (dd, 1H, *J* = 8.4, 2.1 Hz, 6-Hindol), 3.70 (s, 3H, OMe), 3.44 (s, 2H, CH<sub>2</sub>), 2.50 (s, 3H, SMe), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  176.0 (CO<sub>2</sub>H), 168.8 (C=O), 160.8 (C-OMe), 142.5 (C4"arom), 138.9 (C-SMe), 133.4 (C3aindol + C2indol), 132.3, 131.5, 130.6, 129.5, 128.7, 127.0 (Carom ), 114.1 (C3indol + C7indol ), 108.7 (C6indol), 100.9 (C4indol), 55.2 (OMe), 34.2 (CH<sub>2</sub>), 14.9 (SMe), 11.6 (Me). Anal. calcd for C<sub>25</sub>H<sub>23</sub>NO<sub>4</sub>S (445.53): C, 70.09; H, 5.20; N, 3.14. Found: C, 69.85; H, 5.11; N, 2.89.

**2-(5-Methoxy-2-methyl-1-(4'-nitro-[1,1'-biphenyl]4-carbonyl)-indol-3-yl)acetic acid (23):** From 4-nitrophenylboronic acid 11 (47 mg). Yield: 120 mg (81%), as a brown powder, mp 198-200 °C.<sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>): δ 10.42 (s, 1H, CO<sub>2</sub>H), 8.34 (dd, 2H, *J* = 8.1, 2.0 Hz, 3"-H<sub>arom-NO2</sub> + 5"-H<sub>arom-NO2</sub>), 7.89 (dd, 2H, *J* = 8.3, 2.3 Hz, 2'-Harom + 6'-Harom), 7.62-6.95 (m, 4H, 3'-Harom + 5'-Harom +2"-H<sub>arom-NO2</sub> + 6"-H<sub>arom-NO2</sub>), 6.94 (d, 1H, *J* = 8.3 Hz, 7-H<sub>indol</sub>), 6.89 (d, 1H, *J* = 2.2 Hz, 4-H<sub>indol</sub>), 6.54 (dd, 1H, *J* = 2.2, 8.3 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.44 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>): δ 176.1 (CO<sub>2</sub>H), 169.0 (C=O), 160.7 (C-OMe), 147.5 (C1"<sub>arom-NO2</sub>), 146.8 (C-NO<sub>2</sub>), 146.2 (C4'<sub>arom</sub>), 135.9 (C3a<sub>indol</sub>), 132.3 (C2<sub>indol</sub>), 131.4, 130.8, 128.7, 127.3, 124.0 (C<sub>arom</sub>), 114.0 (C3<sub>indol</sub> + C7<sub>indol</sub>), 110.2 (C6<sub>indol</sub>), 100.8 (C4<sub>indol</sub>), 55.2 (OMe), 30.7 (CH2), 11.6 (Me). Anal. calcd for C<sub>25</sub>H<sub>20</sub>N<sub>2</sub>O<sub>6</sub> (444.44): C, 67.56; H, 4.54; N, 6.30. Found: C, 67.32; H, 4.48; N, 6.02.

### 2-(1-(4'-(Ethoxycarbonyl)-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3yl)aceticacid (24):

From 4-ethoxycarbonylphenyl boronic acid 12 (54 mg). Yield: 110 mg (71%), as a yellow product, mp 240-242 °C.<sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  7.95 (dd, 2H, J = 7.8 Hz, 2.1 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.87 (dd, 2H, J = 8.0 Hz, 2.0 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.86 (br s., 2H, 2"-H<sub>arom-OEt</sub> + 6"-H<sub>arom-OEt</sub>), 7.04 (m, 3H, 4-H<sub>indol</sub> + 3"-H<sub>arom</sub> + 5"-H<sub>arom</sub>), 6.93 (d. 1H, J = 8.3 Hz, 7-H<sub>indol</sub>), 6.73 (dd, 1H, J = 8.3, 2.3 Hz, 6-H<sub>indol</sub>), 4.10 (q, 2H, J = 7.1 Hz,  $CH_2CH_3$ ), 3.76 (s, 3H, OMe), 3.34 (s, 2H, CH<sub>2</sub>), 1.20 (t, 3H, CH<sub>2</sub>CH<sub>3</sub>). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  174.3 (CO<sub>2</sub>H), 167.8 (C=O +  $CO_2Et$ ), 157.9 (C-OMe), 155.5 (C-CO<sub>2</sub>Et), 145.2 (C4'<sub>arom</sub>), 137.6 (C3a<sub>indol</sub>), 135.3 (C2<sub>indol</sub>), 134.0 (C1"<sub>arom-CO2Et</sub>), 131.2, 130.5, 130.1, 129.1, 128.5, 127.7 (C<sub>arom</sub>), 114.5 (C3<sub>indol</sub> + C3" + C5"<sub>arom-CO2Et</sub>), 112.7 (C7<sub>indol</sub>), 111.4 (C6<sub>indol</sub>), 101.5 (C4<sub>indol</sub>), 60.3 (OCH<sub>2</sub>CH<sub>3</sub>), 55.3 (OMe), 29.3 (CH<sub>2</sub>), 14.1 (OCH<sub>2</sub>CH<sub>3</sub>), 13.2 (Me). Anal. calcd for C<sub>27</sub>H<sub>25</sub>NO<sub>5</sub> (443.49): C, 73.12; H, 5.68; N, 3.16. Found: C, 72.89; H, 5.58; N, 2.89.

# 2-(1-(4'-Hydroxy)-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (25):

From 4-hydroxyphenylboronic acid 13 (39 mg). Yield: 121 mg (87%), as a brown powder, mp 218-220 °C.<sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$  10.43 (s, 1H, CO<sub>2</sub>H), 7.89 (dd, 2H, *J* = 8.4, 1.8 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.61 (dd, 2H, *J* = 8.1, 2.0 Hz, 2"-H<sub>arom-OH</sub> +6"-H<sub>arom-OH</sub>), 7.31 (dd, 2H, *J* = 8.4, 1.8 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.05 (dd, 2H, *J* = 8.1, 2.0 Hz, 3"-H<sub>arom-OH</sub> + 5"-H<sub>arom-OH</sub>), 6.96 (d, 1H, *J* = 2.5 Hz, 4-H<sub>indol</sub>), 6.82 (d, 1H, *J* = 8.6 Hz, 7-H<sub>indol</sub>), 6.55 (dd, 1H, *J* = 8.6, 2.5 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.45 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  176.0 (CO<sub>2</sub>H), 168.9 (C=O), 161.2 (C4"-OH), 156.7 (C-OMe), 146.8 (C4'<sub>arom</sub>), 138.9 (C3a<sub>indol</sub>), 133.4 (C2<sub>indol</sub>), 132.3, 130.8, 130.1, 129.5, 127.0 (C<sub>arom</sub>), 116.9, 116.6 (C3"<sub>arom-OH</sub> + C5"<sub>arom-OH</sub>), 115.6 (C3<sub>indol</sub> + C7<sub>indol</sub>), 110.2 (C6<sub>indol</sub>), 101.6 (C4<sub>indol</sub>), 55.2 (OMe), 30.8 (CH<sub>2</sub>), 11.6 (Me). Anal. calcd for C<sub>25</sub>H<sub>21</sub>NO<sub>5</sub> (415.44): C, 72.28; H, 5.10; N, 3.37. Found: C, 71.97; H, 4.99; N, 3.12.

# 2-(1-(3'-Cyano-[1,1'-biphenyl]4-carbonyl)-5-methoxy-2-methyl-indol-3-yl)aceticacid (26):

From 3-cyanophenylboronic acid 14 (41 mg). Yield: 119 mg (84%), as a yellow powder, mp 228-230 °C.<sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.41 (s, 1H, CO<sub>2</sub>H), 8.03-7.88 (m, 4H, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub> + 4"-H<sub>arom-CN</sub> + 6"-H<sub>arom-CN</sub>), 7.60 (dd, 2H, *J* = 8.1, 2.1 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.31 (m, 2H, 3"-H<sub>arom-CN</sub> + 5"-H<sub>arom-CN</sub>), 7.05, 7.04 (2xd, 2H, *J* = 2.2 Hz, 7-H<sub>indol</sub> + 4-H<sub>indol</sub>), 6.55 (dd, 1H, *J* = 8.3, 2.2 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.45 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  176.0 (CO<sub>2</sub>H), 168.9 (C=O), 161.6 (C-OMe), 152.4 (C4'<sub>arom</sub>), 144.6 (C1"<sub>arom-CN</sub>), 138.8 (C3a<sub>indol</sub>), 133.4 (C2<sub>indol</sub>), 132.3, 131.4, 130.8, 129.5, 128.7, 126.9 (C<sub>arom</sub>), 117.0 (CN), 112.3 (C3<sub>indol</sub> + C3"<sub>arom</sub>), 110.2 (C7<sub>indol</sub>), 108.6 (C6<sub>indol</sub>), 100.8 (C4<sub>indol</sub>), 55.2 (OMe), 30.7 (CH2), 11.6 (Me). Anal. calcd for C<sub>26</sub>H<sub>20</sub>N<sub>2</sub>O4 (424.45): C, 73.57; H, 4.75; N, 6.60. Found: C, 73.32; H, 4.65; N, 6.41.

### 2-(5-Methoxy-2-mehyl-1-(4'-(trifluoromethyl)-[1,1'-biphenyl]4-carbonyl)-indol-3yl)aceticacid (27):

From 4-trifluoromeyhylphenylboronic acid 15 (53 mg). Yield: 135 mg (88%), as a yellow powder, mp 218-220°C. <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.42 (s, 1H, CO2H), 7.94 (dd, 1H, J = 7.9, 2.1 Hz, 5'-H<sub>arom</sub>), 7.89 (dd, 2H, J = 8.1, 2.2 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.66 (dd, 1H, J = 7.9, 2.1 Hz, 3'-H<sub>arom</sub>), 7.43 (dd, 2H, J = 7.8, 2.5 Hz, 3"-H<sub>arom-CF3</sub> + 5"-H<sub>arom-CF3</sub>), 7.30 (dd, 2H, J = 7.8, 2.5 Hz, 2"-H<sub>arom</sub> + 6"-H<sub>arom</sub>), 6.95 (d, 1H, J = 2.2 Hz, 4-H<sub>indol</sub>), 6.54 (dd, 1H, J = 8.2, 2.1 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.43 (s, 2H, CH<sub>2</sub>), 2.27(s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  176.1 (CO<sub>2</sub>H), 168.9 (C=O), 154.6 (C-OMe), 146.0 (C4'<sub>arom</sub>), 143.9 (C1"<sub>arom-CF3</sub>), 138.9 (C3a<sub>indol</sub>), 133.4 (C2<sub>indol</sub>), 132.3, 130.8, 130.1, 129.5,

127.0, 125.8, 123.0 ( $C_{arom}$ ), 113.2 ( $C3_{indol}$ ), 110.2 ( $C7_{indol}$ ), 100.8 ( $C4_{indol}$ ), 55.2 (OMe), 30.7 (CH2), 11.6 (Me). Anal. calcd for  $C_{26}H_{20}F_3NO_4$  (467.44): C, 66.81; H, 4.31; N, 3.00. Found: C, 66.59; H, 4.27; N, 2.78.

**2-(1-4-(5-Formylthiophen-2-yl)benzoyl)-5-methoxy-2-methyl-indol-3-yl)acetic acid (28):** From 5-formyl-2-thiopheneboronic acid 16 (44 mg). Yield: 129 mg (89%), as a brown powder, mp 208-210°C. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$  10.42 (s, 1H, CO<sub>2</sub>H), 9.80 (s, 1H, CHO), 7.88 (dd, 2H, *J* = 8.1, 2.2 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.64-7.54 (m, 3H, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub> + 5-H<sub>thiophen</sub>), 7.31 (d, 1H, *J* = 6.7 Hz, 4-H<sub>thiophen</sub>), 7.04 (d, 1H, *J* = 8.0 Hz, 7-H<sub>indol</sub>), 6.95 (d, 1H, *J* = 2.1 Hz, 4-H<sub>indol</sub>), 6.54 (dd, 1H, *J* = 8.0, 2.2 Hz, 6-H<sub>indol</sub>), 3.70 (s, 3H, OMe), 3.43 (s, 2H, CH<sub>2</sub>), 2.27 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  182.8 (CHO), 176.0 (CO<sub>2</sub>H), 167.2 (C=O), 152.4 (C-OMe), 148.3 (C2"<sub>thiophen</sub>), 142.1 (C5"<sub>thiophen</sub>), 139.0 (C4'<sub>arom</sub>), 138.2 (C4"<sub>thiophen</sub>), 136.3 (C2<sub>indol</sub>), 133.4, 132.3, 131.4, 130.8, 130.1, 129.5, 128.6 (C<sub>arom</sub>), 114.7 (C(3)<sub>indol</sub>), 110.8 (C6<sub>indol</sub> + C7<sub>indol</sub>), 100.8 (C4<sub>indol</sub>), 55.2 (OMe), 28.7 (CH<sub>2</sub>), 11.6 (Me). Anal. calcd for C<sub>24</sub>H<sub>19</sub>NO<sub>6</sub>S (433.48): C, 66.50; H, 4.42; N, 3.23. Found: C, 66.29; H, 4.35; N, 3.01.

General procedure of preparation of indomethacin having triaryl groups *via* Suzuki-Miyaura cross-coupling reaction (29-31). The compounds were prepared by following the Suzuki reaction procedure described in method A, using instead 2 mol. equiv. of arylboronic acid (0.56 mmol),  $Pd(OAc)_2$  (38 mg, 0.17 mmol) and  $PPh_3$  (132 mg, 0.50 mml).

### 2-(5-Methoxy-2-mehyl-1-(4'-nitro-[1,1'-biphenyl]-4-carbonyl)-indol-3-yl)-1-(4nitrophenyl)ethanone (29):

From 4-nitrophenylboronic acid 11 (94 mg). Yield: 106 mg (69%), as a brown powder, mp 242-245°C. <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta^1$ H NMR (DMSO- $d_6$ ):  $\delta$  8.36-8.27 (m, 6H, H<sub>arom</sub>), 7.87-7.13 (m, 6H, H<sub>arom</sub>), 7.01 (d, 1H, *J* = 8.2 Hz, 7-H<sub>indol</sub>), 6.92 (d, 1H, *J* = 2.3 Hz, 4-H<sub>indol</sub>), 6.60 (dd, 1H, *J* = 2.3, 8.3 Hz, 6-H<sub>indol</sub>), 3.72 (s, 3H, OMe), 3.46 (s, 2H, CH<sub>2</sub>), 2.25 (s, 3H, Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  195.2 (CH<sub>2</sub>CO), 168.5 (C=O), 159.3 (C-OMe), 151.3, 147.1 (2xC-NO<sub>2</sub>), 146.5 (C4'<sub>arom</sub> + C1"<sub>arom</sub>), 140.0 (C<sub>arom</sub>), 136.1 (C3a<sub>indol</sub>), 133.2 (C2<sub>indol</sub>), 131.6, 130.8, 130.4, 129.0, 128.8, 128.5, 127.9, 124.0 (C<sub>arom</sub>), 113.0 (C-6<sub>indol</sub> + C7<sub>indol</sub>), 108.2 (C3<sub>indol</sub>), 100.2 (C4<sub>indol</sub>), 55.6 (OMe), 33.9 (CH2), 12.4 (Me).Anal. calcd for C<sub>31</sub>H<sub>23</sub>N<sub>3</sub>O<sub>7</sub> (549.53): C, 67.75; H, 4.22; N, 7.65. Found: C, 67.54; H, 4.14; N, 7.41.

# 2-(1-(4'-Hydroxy-[1,1'-biphenyl]-4-carbonyl)-5-methoxy1-2-methyl-indol-3-yl)-1-(4 hydroxyphenyl)ethanone (30):

From 4-hydroxyphenylboronic acid 13 (78 mg). Yield: 90 mg (65%), as a pale brown powder, mp 232-235°C. <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>):  $\delta$ 7.91 (dd, 2H, *J* = 8.5, 2.1 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.73 (m, 4H, 2-H<sub>arom-OH</sub> +6-H<sub>arom-OH</sub>), 7.69 (dd, 2H, *J* = 8.1, 2.0 Hz, 2''-H<sub>arom-OH</sub> +6''-H<sub>arom-OH</sub>), 7.31-7.11 (m, 4H, 3-H<sub>arom-OH</sub> +5-H<sub>arom-OH</sub> + 3''-H<sub>arom-OH</sub> +5''-H<sub>arom-OH</sub>), 6.98 (d, 1H, *J* = 8.0 Hz, 7-H<sub>indol</sub>), 6.94 (d, 1H, *J* = 2.2 Hz, 4-H<sub>indol</sub>), 6.63 (dd, 1H, *J* = 8.0, 2.2 Hz, 6-H<sub>indol</sub>), 3.71 (s, 3H, OMe), 3.44 (s, 2H, CH<sub>2</sub>), 2.23 (s, 3H, Me). <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  194.9 (CH<sub>2</sub>CO), 168.2 (C=O), 161.2 (C<sub>arom</sub>-OH), 159.3, 159.1 (C-OMe + C4''<sub>arom</sub>-OH), 145.5 (C4'<sub>arom</sub>), 136.8 (C3a<sub>indol</sub>), 133.9 (C2<sub>indol</sub>), 133.0, 131.6, 130.8, 130.4, 128.8, 128.5, 127.9, 115.8, 114.9 (C<sub>arom</sub>), 112.9 (C-6<sub>indol</sub> + C7<sub>indol</sub>), 106.9 (C3<sub>indol</sub>), 100.1 (C4<sub>indol</sub>), 55.5 (OMe), 34.2 (CH2), 12.7 (Me). Anal. calcd for C<sub>31</sub>H<sub>23</sub>N<sub>3</sub>O<sub>7</sub> (491.18): C, 75.75; H, 5.13; N, 2.85. Found: C, 75.54; H, 5.02; N, 2.69.

### 4'(3-(2-(3-Cyanophenyl)-2-oxoethyl)-5-methoxy-2-methyl-indole-1-carbonyl)-1,1'biphenyl]-3-carbonitrile (31):

From 3-cyanophenylboronic acid 14 (83 mg). Yield: 102 mg (71%), as a pale yellow powder, mp 239-242°C. <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$ 8.24 (d, 1H, J = 2.1 Hz, C2<sub>arom-CN</sub>), 8.11 (m, 1H,

C6<sub>arom-CN</sub>), 8.07-7.46 (m, 10H, H<sub>arom</sub>), 7.12, (d, 1H, J = 8.0 Hz, 7-H<sub>indol</sub>), 6.93 (d, 1H, J = 2.0 Hz, 4-H<sub>indol</sub>), 6.69 (dd, 1H, J = 8.0, 2.0 Hz, 6-H<sub>indol</sub>), 3.75 (s, 3H, OMe), 3.51 (s, 2H, CH<sub>2</sub>), 2.24 (s, 3H, Me).<sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>):  $\delta$  195.6 (CH<sub>2</sub>CO), 168.1 (C=O), 160.8 (C-OMe), 149.0(C4'<sub>arom</sub>), 143.9 (C1"<sub>arom-CN</sub>), 138.2 (C3a<sub>indol</sub>), 136.2 (C1<sub>arom-CN</sub>), 135.1 (C4<sub>arom-CN</sub>), 133.1 (C2<sub>indol</sub>), 132.0, 131.4, 130.5, 130.0, 129.1, 128.5, 128.2, 127.9 (C<sub>arom</sub>), 117.8 (CN), 112.9 (C3"<sub>arom-CN</sub> + C7<sub>indol</sub>), 112.1 (C6<sub>indol</sub> + C3<sub>arom-CN</sub>), 106.4 (C3<sub>indol</sub>), 100.1 (C4<sub>indol</sub>), 55.4 (OMe), 34.5 (CH2), 12.5 (Me). Anal. calcd for C<sub>33</sub>H<sub>23</sub>N<sub>3</sub>O<sub>3</sub> (509.55): C, 77.78; H, 4.55; N, 8.75. Found: C, 77.53; H, 4.41; N, 8.02.

# *Methyl2-(3-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)acetyl)thioureido)carboxylic acid esters (33-35):*

A solution of 4 (200 mg, 0.53 mmol) and NH<sub>4</sub>NCS (40 mg, 0.53 mmol) in dry acetone (15 mL) was heated under reflux for 1h. After cooling the solution was filtered, the desired amino acid esters hydrochloride in dry acetone (15 mL) were added rapidly on the above solution and vigorous stirring and reflux for 6 h (monitored by TLC, eluent: CHCl<sub>3</sub>-MeOH 95:5). After cooling, an excess of crushed ice was poured on the mixture with vigorous stirring. The resulting compound was collected, washed with acetone and recrystallized twice from EtOH.

### *Methyl2-(3-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)acetyl) thioureido)acetate (33):*

From L-alanine methyl ester hydrochloride (74 mg, 0.53 mmol). Yield: 194 mg (75%), as a yellow powder, mp 81-82°C,  $R_f = 0.72$ . <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.82 (s, 1H, NH), 7.82 (dd, 2H, J = 8.1, 1.9 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.54 (dd, 2H, J = 8.1, 1.9 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.36 (d, 1H, J = 8.1 Hz, 7-H<sub>indol</sub>), 7.14 (d, 1H, J = 1.8 Hz, 4-H<sub>indol</sub>), 6.64 (dd, 1H, J = 8.1, 1.8 Hz, 6-H<sub>indol</sub>), 3.83, 3.81 (2xs, 6H, CH<sub>2</sub>-8 + OMe), 3.69 (s, 3H, CO<sub>2</sub>Me), 3.44 (br s., 1H, 13-H), 2.34 (s, 3H, C(2)-Me), 1.23 (s, 3H, C(13)-Me). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  187.0 (C=S), 173.3 (CO<sub>2</sub>Me), 171.0 (C(9)=O), 167.7 (C=O), 155.5 (C-OMe), 139.1 (C-Cl), 137.7 (C3a<sub>indol</sub>), 132.5 (C2<sub>indol</sub>), 130.8, 130.5, 129.8, 128.9, 127.0 (C<sub>arom</sub>), 111.5 (C6<sub>indol</sub> + C7<sub>indol</sub>), 106.4 (C-3), 101.5 (C4<sub>indol</sub>), 58.3 (C-13), 55.7 (OMe), 51.9 (CO<sub>2</sub>Me), 29.4 (C-8), 17.3 (C(13)-Me), 12.8 (C2-Me). Anal. calcd for C<sub>24</sub>H<sub>24</sub>ClN<sub>3</sub>O<sub>5</sub>S (501.98): C, 57.42; H, 4.82; N, 8.37. Found: C, 57.19; H, 4.76; N, 8.11.

#### *Methy2-(3-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetyl) thioureido)butanoate (34):*

From L-valine methyl ester hydrochloride (89 mg, 0.53 mmol). Yield: 244 mg (87%), as a green powder, mp 218-220°C,  $R_f = 0.78$ . <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  10.60 (s, 1H, NH), 10.02 (s, 1H, NH), 7.90 (dd, 2H, J = 8.1, 2.3 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.50 (dd, 2H, J = 8.1, 2.3 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.33 (d, 1H, J = 7.9 Hz, 7-H<sub>indol</sub>), 7.18 (d, 1H, J = 2.0 Hz, 4-H<sub>indol</sub>), 6.67 (dd, 1H, J = 7.9, 2.0 Hz, 6-H<sub>indol</sub>), 3.85 (s, 3H, OMe), 3.74 (s, 2H, CH<sub>2</sub>-8), 3.64 (s, 3H, CO<sub>2</sub>Me), 3.24 (dd, 1H, J = 8.5, 5.0 Hz, 13-H), 2.37 (s, 3H, C(2)-Me), 1.23 (s, 3H, C(13)-Me), 1.84 (m, 1H, 14-H), 1.07 (d, 6H, J = 6.5 Hz, 2xMe).<sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  186.8 (C=S), 172.3 (C(9)=O), 170.5 (CO<sub>2</sub>Me), 167.8 (C=O), 155.3 (C-OMe), 140.3 (C-Cl), 137.6 (C3a<sub>indol</sub>), 135.5 (C2<sub>indol</sub>), 131.1, 130.5, 130.1, 129.1 (C<sub>arom</sub>), 112.8 (C7<sub>indol</sub>), 111.4 (C6<sub>indol</sub>), 107.3 (C-3), 101.6 (C4<sub>indol</sub>), 60.4 (C-13), 55.4 (OMe), 51.1 (CO<sub>2</sub>Me), 30.7 (C-8), 29.3 (C-14), 18.3 (2xMe), 13.2 (C(2)-Me). Anal. calcd for C<sub>26</sub>H<sub>28</sub>ClN<sub>3</sub>O<sub>5</sub>S (530.04): C, 58.92; H, 5.32; N, 7.93. Found: C, 58.73; H, 5.22; N, 7.69.

# *Methyl2-(3-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetyl)thioureido)-3-mercapto propanoate (35):*

From cysteine methyl ester hydrochloride (91 mg, 0.53 mmol). Yield (0.19 g) (95%), as a yellow powder, mp168-170°C,  $R_f = 0.72$ . <sup>1</sup>H NMR (DMSO-d<sub>6</sub>):  $\delta$  9.91 (s, 1H, NH), 9.64 (s,

1H, NH), 7.75 (dd, 2H, J = 7.8, 2.0 Hz, 2'-H<sub>arom</sub> + 6'-H<sub>arom</sub>), 7.56 (dd, 2H, J = 7.8, 2.03 Hz, 3'-H<sub>arom</sub> + 5'-H<sub>arom</sub>), 7.25 (d, 1H, J = 8.0 Hz, 7-H<sub>indol</sub>), 7.21 (d, 1H, J = 2.0 Hz, 4-H<sub>indol</sub>), 6.92 (dd, 1H, J = 8.0, 2.1 Hz, 6-H<sub>indol</sub>), 3.77 (s, 3H, OMe), 3.75 (m, 3H, CH<sub>2</sub>-8 + 13-H), 3.67 (s, 3H, CO<sub>2</sub>Me), 3.46 (m, 2H, CH<sub>2</sub>-SH), 2.27 (s, 3H, C(2)-Me).<sup>13</sup>C NMR (DMSO-d<sub>6</sub>):  $\delta$  186.2 (C=S), 173.0 (C(9)=O), 171.9 (CO<sub>2</sub>Me), 167.1 (C=O), 151.1 (C-OMe), 139.3 (C-Cl), 136.6 (C3a<sub>indol</sub>), 136.2 (C2<sub>indol</sub>), 132.4, 130.9, 129.4, 129.2, 128.4 (C<sub>arom</sub>), 112.8 (C7<sub>indol</sub>), 112.5 (C7<sub>indol</sub>), 110.6 (C6<sub>indol</sub>), 109.3 (C-3), 99.4 (C4<sub>indol</sub>), 69.6 (C-13), 55.3 (OMe), 51.7 (CO<sub>2</sub>Me), 30.6 (C-8), 26.4 (CH<sub>2</sub>-SH), 13.2 (C2-Me). Anal. calcd for C<sub>24</sub>H<sub>24</sub>ClN<sub>3</sub>O<sub>5</sub> (535.05): C, 53.98; H, 4.43; N, 7.87. Found: C, 53.77; H, 4.32; N, 7.58.

General procedure for the preparation of methyl 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1*H*-indol-3-yl)acetamido carboxylate derivatives (36-41). To a cold solution of amino acid esters hydrochloride (0.28 mmol) at - 5°C in MeCN (20 mL), indomethacin 1 (100 mg, 0.28 mmol), HOBT (38 mg, 0.28 mmol) and DCC (58 mg, 0.28 mmol) were added successively. The reaction mixture was stirred at 0°C for 1 h, 5°C for 1h, and 23°C for 16 h. Dicyclohexylurea (DCU) was filtered and the filterate was evaporated to dryness. The residue was stirred in ethyl acetate (20 mL), filtered, washed successively with saturated (NaCl) solution (20 mL), 5% NaHCO<sub>3</sub> solution (20 mL), 1.0 mol L<sup>-1</sup> HCI (10 mL), and finally washed with water. The organic layer was dried by (Na<sub>2</sub>SO<sub>4</sub>), filtered, and the filtrated was evaporated to dryness and recrystallized from EtOH.

## Methyl2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamido acetate (36):

From glycine methyl ester hydrochloride (35 mg). Yield: 84 mg (70%), as a brown powder, mp 68-70°C,  $R_{\rm f}$  = 0.48. <sup>1</sup>H NMR (DMSO- $d_{\rm 6}$ ):  $\delta$  8.50 (d, 1H,  $J_{\rm NH,H11}$  = 5.5 Hz, NH), 7.71-7.62 (m, 4H, H<sub>arom</sub>), 7.15 (d, 1H, J = 8.0 Hz, 7-H<sub>indol</sub>), 6.94 (br s., 2H, J = 2.0 Hz, 4-H<sub>indol</sub> + 6-H<sub>indol</sub>), 3.78 (d, 2H, J = 2.2 Hz, CH<sub>2</sub>-11), 3.75 (br s., 5H, CH<sub>2</sub>-8 + OMe), 3.65 (s, 3H, CO<sub>2</sub>Me), 2.23 (s, 3H, C(2)-Me). <sup>13</sup>C NMR (DMSO- $d_{\rm 6}$ ):  $\delta$ 173.0 ( $CO_{2}$ Me), 170.1 (C(9)=O), 167.7 (C=O), 156.6 (C-OMe), 137.1 (C-Cl), 135.2 (C3a<sub>indol</sub> + C-2), 131.1, 130.9, 129.0 (C<sub>arom</sub>), 113.8 (C7<sub>indol</sub>), 110.6 (C6<sub>indol</sub>), 106.1 (C-3), 101.9 (C4<sub>indol</sub>), 55.5 (OMe), 53.6 (CO<sub>2</sub>Me), 43.5 (C-11), 30.7 (C-8), 14.0 (C2-Me).Anal. calcd for C<sub>22</sub>H<sub>21</sub>CIN<sub>2</sub>O<sub>5</sub> (428.87): C, 61.61; H, 4.94; N, 6.53. Found: C, 61.48; H, 4.79; N, 6.33.

**Methyl2-(1-(4-chorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamidopropanoate(37):** From L-alanine methyl ester hydrochloride (39 mg): Yield: 86 mg (65%), as a semi-solid,  $R_{\rm f}$ = 0.68. <sup>1</sup>H NMR (DMSO- $d_{\rm 6}$ ): δ 8.53 (d, 1H,  $J_{\rm NH,H11}$  = 5.8 Hz, NH), 7.66 (br s., 4H, H<sub>arom</sub>), 7.08 (d, 1H, J = 7.9 Hz, 7-H<sub>indol</sub>), 6.94 (br s., 2H, 4-H<sub>indol</sub> + 6-H<sub>indol</sub>), 4.18 (dd, 2H, J = 7.2, 5.8 Hz, 11-H), 3.75 (s, 5H, CO<sub>2</sub>Me + OMe), 3.65 (s, 3H, CH<sub>2</sub>-8), 2.30 (s, 3H, C2-Me), 1.70 (d, 3H, J= 7.1 Hz, C11-Me). <sup>13</sup>C NMR (DMSO- $d_{\rm 6}$ ): δ173.2 (CO<sub>2</sub>Me), 171.6 (C9=O), 167.7 (C=O), 156.7 (C-OMe), 137.2 (C-Cl), 134.6 (C3a<sub>indol</sub>), 133.2 (C-2), 131.9, 130.9, 128.9 (C<sub>arom</sub>), 112.6 (C7<sub>indol</sub>), 110.7 (C(6)<sub>indol</sub>), 105.5 (C-3), 102.4 (C4<sub>indol</sub>), 57.5 (OMe), 55.3 (C-11), 53.2 (CO<sub>2</sub>Me), 29.9 (C-8), 18.2 (C11-Me), 13.3 (C2-Me).Anal. calcd for C<sub>23</sub>H<sub>23</sub>ClN<sub>2</sub>O<sub>5</sub> (442.89): C, 62.37; H, 5.23; N, 6.33. Found: C, 62.09; H, 5.11; N, 6.20.

# *Methyl2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamido)-3-hydroxy)propanoate (38):*

From L-serine methyl ester hydrochloride (43 mg). Yield: 104 mg (78%), as a brown powder, mp 78-80°C,  $R_{\rm f}$  = 0.50. <sup>1</sup>H NMR (DMSO- $d_{\rm 6}$ ):  $\delta$  8.38 (br s., 1H, NH), 7.64 (br s., 4H, H<sub>arom</sub>), 7.06 (br s., 2H,4-H<sub>indol</sub> + 7-H<sub>indol</sub>), 6.93 (d., 1H, *J* = 7.8 Hz, 6-H<sub>indol</sub>), 4.20 (dd, 1H, *J* = 8.3, 4.8 Hz, 11-H), 3.78 (s, 5H, CO<sub>2</sub>*M*e), 3.59 (s, 3H, OMe), 3.57 (s, 2H, CH<sub>2</sub>-8), 3.58 (t, 1H, *J* = 5.7 Hz, OH), 3.40 (s, 3H, CH<sub>2</sub>OH), 2.31 (s, 3H, C(2)-Me). <sup>13</sup>C NMR (DMSO- $d_{\rm 6}$ ):  $\delta$ 172.4 (CO<sub>2</sub>Me), 170.2 (C9=O), 167.8 (C=O), 156.7 (C-OMe), 137.5 (C-Cl), 135.1(C3a<sub>indol</sub> + C-2),

131.7, 131.1, 130.1, 129.0 ( $C_{arom}$ ), 112.4 ( $C7_{indol}$ ), 111.5 ( $C6_{indol}$ ), 104.4 (C-3), 102.0 ( $C4_{indol}$ ), 59.7 (C-11), 55.4 (OMe), 53.5 ( $CO_2Me$ ), 31.3 (C-8), 26.2 (CH<sub>2</sub>-OH), 13.9 (C2-*Me*).Anal. calcd for  $C_{23}H_{23}CIN_2O_5S$  (474.96): C, 58.16; H, 4.88; N, 5.90. Found: C, 57.92; H, 4.69; N, 5.69.

# *Methyl2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamido)-3-methyl butanoate(39):*

From L-valine methyl ester hydrochloride (47 mg). Yield: 94 mg (71%), as a light-yellow powder, mp 110-113°C,  $R_f = 0.63$ . <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  8.61 (d, 1H,  $J_{NH,H11} = 5.6$  Hz, NH), 7.69 (br s., 4H, H<sub>arom</sub>), 7.12 (d, 1H, J = 8.1 Hz, 7-H<sub>indol</sub>), 7.01 (br s., 2H, 4-H<sub>indol</sub> + 6-H<sub>indol</sub>), 4.20 (dd, 2H, J = 7.4, 5.6 Hz, 11-H), 3.77 (s, 5H, CO<sub>2</sub>Me), 3.72 (s, 3H, OMe), 3.65 (s, 3H, CH<sub>2</sub>-8), 2.28 (s, 3H, C(2)-Me), 2.07 (m, 1H, 12-H), 0.81, 0.83 (2s, 6H, 2xMe). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$ 172.8 (CO<sub>2</sub>Me), 170.9 (C(9)=O), 167.1 (C=O), 156.5 (C-OMe), 137.0 (C-CI), 134.7 (C3a<sub>indol</sub>), 133.5 (C-2), 131.6, 130.3, 128.6, 128.3 (C<sub>arom</sub>), 112.8 (C7<sub>indol</sub>), 110.5 (C6<sub>indol</sub>), 105.3 (C-3), 102.5 (C4<sub>indol</sub>), 57.5 (OMe), 62.1 (C-11), 52.4 (CO<sub>2</sub>Me), 31.1 (C-8), 30.5 (C-12), 18.9, 17.6 (2xMe), 13.1 (C2-Me).Anal. calcd for C<sub>25</sub>H<sub>27</sub>ClN<sub>2</sub>O<sub>5</sub> (470.95): C, 63.76; H, 5.78; N, 5.95. Found: C, 63.53; H, 5.69; N, 5.81.

# *Methyl2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamido)-4-methyl pentanoate (40):*

From L-leucine methyl ester hydrochloride (51 mg). Yield: 87 mg (64%), as a colorless powder, mp 131-134°C,  $R_f = 0.68$ . <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  8.63 (d, 1H,  $J_{NH,H11} = 5.5$  Hz, NH), 7.70 (br s., 4H, H<sub>arom</sub>), 7.12 (d, 1H, J = 7.9 Hz, 7-H<sub>indol</sub>), 6.98 (br s., 2H, 4-H<sub>indol</sub> + 6-H<sub>indol</sub>), 4.26 (dd, 2H, J = 7.5, 5.5 Hz, 11-H), 3.78 (s, 3H, CO<sub>2</sub>Me), 2.74 (s, 3H, OMe), 4.41 (m, 1H, 11-H), 3.70 (s, 3H, CH<sub>2</sub>-8), 2.05 (m, 2H, CH<sub>2</sub>-12), 1.17 (m, 1H, 13-H), 0.80, 0.82 (2s, 6H, 2xMe). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$ 173.0 ( $CO_2$ Me), 171.2 (C9=O), 167.5 (C=O), 156.5 (C-OMe), 136.9 (C-Cl), 134.2 ( $C3a_{indol}$ ), 133.0 (C-2), 131.7, 130.3, 129.9, 128.1 ( $C_{arom}$ ), 112.9 ( $C7_{indol}$ ), 110.5 ( $C6_{indol}$ ), 105.2 (C-3), 102.1 ( $C4_{indol}$ ), 57.3 (OMe), 56.1 (C-11), 53.8 ( $CO_2Me$ ), 51.1 (C-12), 32.0 (C-8), 30.8 (C-13), 13.1 (C2-Me).Anal. calcd for  $C_{26}H_{29}CIN_2O_5$  (484.97): C, 64.39; H, 6.03; N, 5.78. Found: C, 64.18; H, 5.97; N, 5.62.

## *Methyl6-amino-2-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetamido)-3-hexanoate (41):*

From L-lysine methyl ester hydrochloride (55 mg, 0.28 mmol). Yield: 111 mg (79%), as a light yellow powder, mp98-102°C,  $R_f = 0.28$ . <sup>1</sup>H NMR (DMSO- $d_6$ ):  $\delta$  8.50 (d, 1H, J = 5.7 Hz, NH), 7.72-7.64 ((m, 4H, H<sub>arom</sub>), 7.08 (d., 1H, J = 7.5 Hz, 7-H<sub>indol</sub>), 6.96, 6.95 (2xbr s., 2H, 4-H<sub>indol</sub> + 6-H<sub>indol</sub>), 5.96 (d, 2H, J = 8.1 Hz, NH<sub>2</sub>), 4.50 (dd, 1H, J = 8.2, 5.7 Hz, 11-H), 3.78 (s, 3H, OMe), 3.75 (s, 3H, CO<sub>2</sub>Me), 3.54 (s, 2H, CH<sub>2</sub>-8), 2.51 (m., 2H, CH<sub>2</sub>-15), 2.23 (s, 3H, C(2)-Me), 1.71 (m, 4H, CH<sub>2</sub>-12 + CH<sub>2</sub>-14), 1.60 (m, 2H, CH<sub>2</sub>-13). <sup>13</sup>C NMR (DMSO- $d_6$ ):  $\delta$  173.0 (CO<sub>2</sub>Me), 170.0 (C9=O), 167.8 (C=O), 156.6 (C-OMe), 137.5 (C-Cl), 135.2 (C3a<sub>indol</sub>), 134.2 (C-2), 132.0,130.9, 130.8, 128.9 (C<sub>arom</sub>), 112.6 (C7<sub>indol</sub>), 110.6 (C6<sub>indol</sub>), 102.5 (C-3), 101.9 (C4<sub>indol</sub>), 56.2 (C-11), 55.4 (OMe), 53.1 (CO<sub>2</sub>Me), 47.5 (C-15),33.3 (C-8), 30.7 (C-12), 28.9 (C-14), 24.4 (C-13), 14.0 (C2-Me). Anal. calcd for C<sub>26</sub>H<sub>30</sub>ClN<sub>3</sub>O<sub>5</sub> (499.99): C, 62.46; H, 6.05; N, 8.40. Found: C, 62.27; H, 5.96; N, 8.25.

## 2.3 Biological Activity

### 2.3.1 Antiviral assay

#### 2.3.1.1 In vitro anti-HIV assay

Evaluation of the antiviral activity of the new synthesized compounds against HIV-1 strain III<sub>B</sub> and HIV-2 strain (ROD) in MT-4 cells was performed using the MTT assay as previously described [28,29]. Briefly, stock solutions (10 x final concentration) of test compounds were added in 25  $\mu$ L volumes to two series of triplicate wells so as to allow simultaneous evaluation of their effects on mock-and HIV-infected cells at the beginning of each experiment. Serial 5-fold dilutions of test compounds were made directly in flat-bottomed 96well microtiter trays using a Biomek 3000 robot (Beckman instruments). Untreated control HIV-and mock-infected cell samples were included for each sample.HIV-1(III<sub>B</sub>) [30] or HIV-2 (ROD) [31] stock (50  $\mu$ L) at 100-300 CCID<sub>50</sub> (50% cell culture infectious dose) or culture medium was added to either the infected or mock-infected wells of the microtiter tray. Mockinfected cells were used to evaluate the effect of test compound on uninfected cells in order to assess the cytotoxicity of the test compound. Exponentially growing MT-4 cells [32] were centrifuged for 5 min at 1,000 rpm and the supernatant was discarded. The MT-4 cells were resuspended at 6 x 10<sup>5</sup> cells/mL and 50  $\mu$ L volumes were transferred to the microtiter tray wells. Five days after infection at ambient temperature, the viability of mock-and HIV-infected cells was examined spectrophotometrically by the MTT assay.

### 2.3.1.2 Anti-HCV assay

Huh 5.2 cells, containing the hepatitis C virus genotype 1b I389luc-ubi-neo/NS3-3'/5.1 replicon [33] were sub-cultured in DMEM supplemented with 10% FCS, 1% non-essential amino acids, 1% penicillin/streptomycin and 2% Geneticin at a ratio of 1:3 to 1:4, and grown for 3 - 4 days in 75 cm<sup>2</sup> tissue culture flasks. One day before addition of the compound, cells were harvested and seeded in assay medium (DMEM, 10% FCS, 1% non-essential amino acids, 1% penicillin/streptomycin) at a density of 6500 cells/well (100  $\mu$ L/well) in 96-well tissue culture microtiter plates for evaluation of anti-metabolic effect and Cultur-Plate (Perkin Elmer) for evaluation of the antiviral effect. The microtiter plates were incubated overnight (37°C, 5% CO<sub>2</sub>, 95-99% relative humidity), yielding a non-confluent cell monolayer.

The evaluation of the anti-metabolic as well as antiviral effect of each compound was performed in parallel. Four-step, 1-to-5 compound dilution series were prepared for the first screen, to collect data for a more detailed dose-response curve, an eight-step, 1-to-2 dilution series was used. Following assay setup, the microtiter plates were incubated for 72 hours (37°C, 5% CO<sub>2</sub>, 95-99% relative humidity). For the evaluation of anti-metabolic effects, the assay medium was aspirated, replaced with 75  $\mu$ L of a 5% MTS solution in phenol red-free medium and incubated for 1.5 hours (37°C, 5% CO<sub>2</sub>, 95-99% relative humidity). Absorbance was measured at a wavelength of 498 nm (Safire<sup>2</sup>, Tecan), and optical densities (OD values) were converted to percentage of untreated controls. For the evaluation of antiviral effects, assay medium was aspirated and the cell monolayers were washed with phosphate-buffered saline (PBS). The wash buffer was aspirated, and 25  $\mu$ L of GloLysis Buffer (Promega) was added allowing for cell lysis to proceed for 5 min at room temperature. Subsequently, 50  $\mu$ L of Luciferase Assay System (Promega) was added, and the luciferase luminescence signal was quantified immediately (1000 ms integration time/well, Safire<sup>2</sup>, Tecan). Relative luminescence units were converted into percentage of untreated controls.

The EC<sub>50</sub> and EC<sub>90</sub> (values calculated from the dose-response curve) represent the concentrations at which 50% and 90% inhibition, respectively, of viral replication is achieved. The CC<sub>50</sub> (value calculated from the dose-response curve) represents the concentration at which the metabolic activity of the cells is reduced by 50% as compared to untreated cells.

A concentration of compound is considered to elicit a genuine antiviral effect in the HCV replicon system when the anti-replicon effect is well above the 70% threshold at concentrations where no significant anti-metabolic activity is observed [33].

### 2.3.1.3 Cells and HCV viruses

The Huh-5-2 and Huh 9-13 HCV subgenomic replicon-containing cells were provided by Prof. R. Bartenschlager (University of Heidelberg, Heidelberg, Germany).

### 2.3.2 Anti-Kinesin Eg5 assay

The ATPase activity of the Eg5 motor domain was measured using the malachite green assay as described earlier [45]. The reactions were performed in reaction buffer (80 mM Pipes, pH 6.8; 1 mMEGTA, 1 mM MgCl<sub>2</sub>, 0.1 mg/mL<sup>-1</sup> BSA, 1 mMtaxol) supplemented with Eg5 (48 nM) fusion protein and microtubules (200 nM). Ten min after compound addition, reactions were started by the addition of ATP (50 mM) and incubated at RT for 7 min. The reactions were stopped by adding perchloric acid (444 mM, Fluka), and the colour reaction was started by adding the developer solution (1M HCI (Sigma), 33 mM malachite green (Sigma), 775 mM ammonium molybdatetetrahydrate (Sigma)). After 20 min, the absorbance at 610 nm was measured using a plate reader (Victor 2, Perkin-Elmer). The IC<sub>50</sub> values were determined in three independent experiments for each compound. Fig. 1 shows the ATPase activity screening assay.



Fig. 1. ATPase activity screening assay

## 3. RESULTS AND DISCUSSION

## 3.1 Chemistry

The Suzuki coupling reaction [34] has been employed in the preparation of new indomethacin analogues. Thus, treatment of indomethacin 1 with the appropriate arylboronic acids (e.g.: 2-fluorophenyl-, 3-fluorophenyl-, 4-fluorophenyl-, 3,4-difluorophenyl-, 3,4-

dimethoxy phenyl-, 2-methylthiophenyl-, 4-nitrophenyl-, 4-ethoxycarbonylphenyl-, 4hydroxyphenyl-, 3-cyanophenyl-, and 4-triflouromethylphenyl boronic acids 5-15 and 5formyl-2-thiopheneboronic acid 16, using a mixture of palladium acetate / triphenylphosphine complex and sodium bicarbonate as a catalyst afforded 17-28 in 72-78% yield, respectively. The N-heterocyclic carbenecomplexs (Pd-NHC) performed Suzuki cross-coupling reaction at reasonable temperature. where the catalyst palladium 1,3-bis(2,6diisopropylphenyl)imidazol-2-ylidene(1,4-naphthoquinone)palladium(0)dimer [Pd(iPr)(NQ)]2 is considered as one of these complexes, exhibited impressive activity in the Suzuki-Miyaura coupling of aryl chlorides with phenyl boronic acids. Accordingly, 1 was treated with aryl boronic acids 5-7 in the presence of 0.5 mole of [Pd(iPr)(NQ)]<sub>2</sub> [35], potassium tert-butoxide in 2-propanol as a solvent. This method afforded almost better yields of 17-19 (85, 83 and 89%, respectively) (Scheme 1). Interestingly, the triaryl-indomethacin analogues 39-31 were obtained in 69, 65 and 71% yield, respectively, from treatment of 4 [15], prepared from chlorination of 1, and 2 mol. equiv. of arylboronic acids (11, 13 and 14), employing Suzuki-Miyaura cross-coupling reaction (Scheme 1).

The structures of 17-28 were identified by the <sup>1</sup>H and <sup>13</sup>C NMR spectra, which are in agreement with the suggested structures, and showed rather similar patterns for the indole scaffold. The 1H NMR spectra were and characterized by the presence of additional aromatic proton and carbon atoms, indicative for arylation of indomethacin 1. The low field singlets at the regions  $\delta$  10.43-10.41 ppm were assigned to CO<sub>2</sub>H signals. The aromatic protons were appeared as doublets, doublet of doublets, multiplets and broad singlets at the regions  $\delta$  8.34-6.55 ppm, whereas H-4 of the indole backbone resonated as doublets at the regions  $\delta$  7.05-6.89 ppm (J ~ 2.2 Hz). 6-H and 7-H protons were appeared as doublet of doublets and doublets at the regions  $\delta$  6.78-6.53 and  $\delta$  7.06-6.82 ppm (J ~ 8.2, 2.2 Hz). respectively. The singlets at the regions  $\delta$  3.47-3.25 ppm were assigned to the methylene protons (CH<sub>2</sub>-8). The other protons of the prepared compounds were fully analyzed (cf. Experimental section). In the <sup>13</sup>C NMR spectra of 17-28, the CO<sub>2</sub>H signals appeared at the regions  $\delta$  176.1-174.3 ppm, while the resonances at  $\delta$  169.0-167.2 ppm were assigned to C=O carbon atoms. The C-OMe(C-5) carbons appeared at the regions  $\delta$  161.6-152.4 ppm, and C-3a resonated at the regions  $\delta$ 139.0-133.3 ppm. C-2 and C-3carbons resonated at the regions  $\delta$  136.3-132.2 ppm, and  $\delta$  116.0-108.7 ppm, respectively, while the resonances at the regions  $\delta$ 101.6-100.0 ppm,  $\delta$  111.4-108.3 ppm, and  $\delta$ 115.4-110.2 ppm were assigned to C-4, C-6 and C-7, respectively. Aromatic carbon atoms and carbons of the other substituents were fully assigned (cf. Experimental section). The structures of 29-31 were identified from their <sup>1</sup>H and <sup>13</sup>C NMR spectra, which almost similar for those of the analogues 23, 25 and 26. In the <sup>13</sup>C NMR of 29-31, the resonances at  $\delta$  195.2, 194.9 and 195.6 ppm were assigned to the carbonyl carbon atoms (CH<sub>2</sub>COAr). The protons and carbon atoms of these compounds were fully analyzed (cf. Experimental section).

Our work was modified by selecting 4, as a precursor for the synthesis of new indomethacinthioureido amino acid analogues to examine their antiviral activity in comparison to 16-27. Thus, treatment of 4 with ammonium thiocyanate in acetone by following Kabbani approach [36]afforded 32,which was directly treated with the amino acid derivatives (I-alanine, I-valine and I-serine methyl esters hydrochloride) to give, after purification, the thioureido derivatives 33-35 in 75, 87, and 95 % yield, respectively. The synthetic reactions are summarized in scheme 2.

American Chemical Science Journal, 4(4): 516-536, 2014



Scheme 1. Condition and reagents: (i) Pd(OAc)<sub>2</sub>, Ph<sub>3</sub>P, K<sub>2</sub>CO<sub>3</sub>, *n*-PrOH, reflux; or (ii) [Pd(iPr)(NQ)]<sub>2</sub> complex, K <sup>tert</sup>OBu, 2-PrOH, 50°C, 1 h



#### Scheme 2. Synthesis of methyl 2-(3-(2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl)acetyl)thioureido)-carboxylate derivatives

The structures of 33-35 were determined from their NMR (<sup>1</sup>H, <sup>13</sup>C), since they showed a similar pattern of the indol moiety and the aromatic protons (*cf.* Experimental Section). CH<sub>2</sub>-8 protons appeared as singlets at  $\delta$  3.83, 3.74 and 3.75 ppm, respectively, while H-13 protons signals were resonated at broad singlet, doublet of doublets (*J* = 8.5, 5.0 Hz) and multiplet at  $\delta$  3.44, 3.24 and 3.75 ppm, respectively. The other protons of amino acid esters were fully analyzed. The <sup>13</sup>C NMR spectra of 33-35 contained almost similar resonance signals of the indole, aromatic and thioureido carbon atoms. The chemical shifts at  $\delta$  187.0, 186.8 and 186.2 ppm were assigned to C=S carbon atoms of the thioureido moiety (C-11), while the resonances at  $\delta$ 173.3, 170.5 and 171.9 ppm were assigned to the carbonyl groups of the

 $CO_2$ Meresidues, respectively. The carbonyl carbon atoms (C-9) appeared at  $\delta 171.0$ , 172.3 and 173.0 ppm, respectively, while the signals oriented at  $\delta \sigma$  155.5, 155.3 and 151.1 ppm were attributed to C-OMe carbon atoms. C-8 carbon atoms were resonated at  $\delta 29.9$ , 30.7 and 30.6 ppm, and C-13 at  $\delta 58.3$ , 60.4 and 69.6 ppm, respectively. The proton spin system of 31 was further identified from DFQ-COSY [37] spectrum, where the doublet of doublets of H<sub>valin</sub>-13at  $\delta_H$  3.24 ppm was found to correlate with C-13 carbon atom at  $\delta_C$  60.4 ppm.

Next, indomethacin 1 was treated with various amino acid methyl esters (glycine, I-alanine, I-serine, I-valine, I-leucine and I-lysine acetates hydrochloride) in the presence of 1-hydroxybenzotriazole (HOBT) and dicyclohexylcarbodiimide (DCC) as coupling reagents to give 36-41 in 64% -79% yield (Scheme 3).

The structures of 36-41 were determined by their <sup>1</sup>H, and <sup>13</sup>C NMR, which showed a similar pattern of indole and aromatic protons. The low field doublets at the regions  $\delta$  8.63-8.38 ppm were assigned to NH group ( $J_{NH,H11} \sim 5.5$  Hz). CH<sub>2</sub>-8 protons appeared as singlets at the regions  $\delta$  4.41-3.57 ppm, while H-11 protons of the amino acid residues (CONHCH) resonated as doublet or doublet of doublets at  $\delta$  4.50-3.78 ppm ( $J_{\text{NH,H11}} \sim 5.5$  Hz for doublet, and ~ 8.0 Hz, ~ 5.5 Hz for doublet of doublets).  $CH_2$ -11 of glycine residue of 36 appeared as doublet at  $\delta$  3.78 Hz (J = 2.2 Hz), while H-11 protons of 37-41 were resonated as doublet of doublets or multiplet at the regions  $\delta$  4.50-4.18 (J ~ 7.9, 5.6 Hz). The other protons of aromatic, indole and amino acid moieties were fully analyzed (cf. Experimental section). In the <sup>13</sup>C NMR spectra of 37-41, the carbon atoms of the carboxylate groups resonated at the regions  $\delta$  173.2-172.4 ppm, while the carbonyl carbon atoms (C9=O) of the amide moiety resonated at the regions  $\delta$  171.6-170.0 ppm. The resonances at the regions  $\delta$  33.3-29.9 ppm were attributed to the carbon atoms of C-8, while the resonances at the regions  $\delta$  62.1-55.3 ppm were assigned to C-11 of the amino acid residues, except for 36 which appeared at  $\delta$ 43.5 ppm. The CHMe, CH<sub>2</sub>OH carbon signals of 37 and 38 appeared at  $\delta$  18.2 and 26.2 ppm, respectively, while C-12 - C-15 of the l-lysine residue were resonated at  $\delta$  30.7, 24.4, 28.9 and 47.7 ppm, respectively. Compound 41 has been selected for further NMR study. Thus, a gradient selected HMBC spectrum [38] of 41 allowed the identification of CH<sub>2</sub>-8of the amide group at  $\delta_{\rm H}$  3.54 ppm from the  $^2J_{C,H}$  correlations to C-3a of the indole ring at  $\delta_{\rm C}$  135.2 ppm and carbonyl carbon atom (C-9) of I-lysine residue at  $\delta_{C}$  170.0 ppm. Similarly, H-11 proton at  $\delta_{H}$  4.50 ppm was identified from it's a  ${}^{3}J_{C,H}$  correlations to C-9 at  $\delta_{X}$  170.0 ppm and C-13 at  $\delta_{\rm C}$  24.4 ppm, as well as two  $^2J_{\rm C,H}$  correlations: one with the carbonyl atom of the acetate group at  $\delta_C$  173.0 ppm and the other with C-12 at  $\delta_C$  30.7 ppm. C-14 at  $\delta_C$  28.9 ppm showed a  ${}^{2}J_{C,H}$  correlation with CH<sub>2</sub>-13 at  $\delta_{H}$  1.60 ppm, while 4-H of the indole ring at  $\delta_{H}$  6.96 ppm showed the same correlation with C-3a at  $\delta_C$  135.2 ppm. Additionally, The carbon atom (C-5) of the indole ring at  $\delta_{\rm C}$  156.6 ppm showed a  ${}^{3}J_{\rm C,H}$  correlation with 7-H at  $\delta_{\rm H}$  7.08 ppm (Fig. 2). Additionally, all the structures of the new synthesized indomethacin analogues have been identified by the <sup>1</sup>H, <sup>13</sup>C HSQC NMR spectroscopy [39].

American Chemical Science Journal, 4(4): 516-536, 2014



Scheme 3. Synthesis of alkyl 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-indol-3-yl) acetamido carboxylate derivatives



Fig. 2.  $J_{C,H}$  correlations in the HMBC NMR spectrum of 41

## 3.2 Bioactivities

## 3.2.1 In-vitro anti-HIV activity

Compounds 17-28, 29-30 and 33-41 were tested for their *In vitro* anti-HIV-1 (strain III<sub>B</sub>) and HIV-2 (strain ROD) activity in human (MT-4) cells based on an MTT assay [28,29]. The results are summarized in Table 1, in which the data for nevirapine (BOE/BIRG587) [40]and azidothymidine (DDN/AZT) [41] were included for comparison purposes. Compound-induced cytotoxicity was also measured in MT-4 cells parallel with the antiviral activity.

Entry	HIV-1 (III <sub>B</sub> )	HIV-2 (ROD)	CC <sub>50</sub>	SI <sup>e</sup>	SI <sup>e</sup>
-	IC <sub>50</sub> (μM) <sup>c</sup>	IC <sub>50</sub> (μM) <sup>c</sup>	(µM) <sup>d</sup>	(III <sub>B</sub> )	(ROD)
4	>32.70	>32.70	32.70	<1	<1
17	>106.00	>106.00	≥106.00	<orx1< td=""><td><orx1< td=""></orx1<></td></orx1<>	<orx1< td=""></orx1<>
18	>99.53	>99.53	99.53	<1	<1
19	> 90.90	>90.90	≥90.90	<orx1< td=""><td><orx1< td=""></orx1<></td></orx1<>	<orx1< td=""></orx1<>
20	>100.00	>100.00	≥ 100.00	<orx1< td=""><td><orx1< td=""></orx1<></td></orx1<>	<orx1< td=""></orx1<>
21	>86.70	>86.70	86.70	<1	<1
22	>104.00	>104.00	≥ 104.00	< orX1	< orX1
23	>67.25	>67.25	67.25	<1	<1
24	>92.68	>92.68	92.68	<1	<1
25	>77.88	>77.88	77.88	<1	<1
26	>103.75	>103.75	103.75	<1	<1
27	>125.00	>125.00	125.00	<1	<1
28	>101.13	>101.13	101.13	<1	<1
29	>12.33	>12.33	12.33	<1	<1
30	>8.83	>8.83	8.83	<1	<1
31	>1.81	>1.81	3.31	6	<1
33	>2.23	>2.23	2.23	<1	<1
34	>11.10	>11.10	11.10	<1	<1
35	>11.80	>11.80	11.80	<1	<1
36	>2.13	>2.13	2.13	<1	<1
37	>10.28	>10.28	10.28	<1	<1
38	>2.06	>2.06	2.06	<1	<1
39	>3.21	>28.89	28.89	9	<1
40	>7.27	>7.27	7.27	<1	<1
41	>1.98	>1.98	1.98	<1	<1
Nevirapine	0.050	>4.00	> 4.00	>80	<1
AZT	0.0022	0.00094	>25	>11363	>26596

Table 1. In-vitro anti-HIV-1 <sup>a</sup> and HIV-2 <sup>b</sup> activity and cytotoxicity of some new
indomethacin analogues

<sup>a</sup>Anti-HIV-1 activity measured with strain III<sub>B</sub>;<sup>b</sup> anti-HIV-2 activity measured with strain ROD; <sup>c</sup> compound concentration required to achieve 50% protection of MT-4 cells from the HIV-1 and HIV-2induced cytopathic effect; <sup>d</sup> compound concentrationthat reduces the viability of mock-infected MT-4 cells by 50 %; <sup>e</sup> SI: selectivity index (CC<sub>50</sub>/IC<sub>50</sub>)

None of these derivatives showed anti-HIV activity, except 31 and 39 with  $IC_{50}$  values of >1.81 and > 3.21 µM ( $CC_{50}$  of 3.31 and 28.89µM), resulting in selectivity index (SI) of 6 and 9, respectively. Derivatives 33, 36, 38 and 41 demonstatred low $CC_{50}$  values of >2.15, >3.03, >2.29, and >1.63 µM, respectively, at concentration of 100 µM, in comparison to the other analogues. However, the above data suggested that the aryInitril as a keto moiety (e.g.: 31) considerably increased the anti-HIV activity, in comparison to the effectiveness of other functional groups and to the other anti-HIV drug having nitrile residue such as rilpivirine [42].

## 3.2.2 In-vitro anti-HCV activity

Compounds 28, 31 and 32 were selected for evaluation of their *In vitro* selective antihepatitis C virus (HCV) activity in the Huh 5-2 replicon system (type 1b, Con1 strain) [33]. The compounds showed EC<sub>50</sub> of 9.56, 8.27 and 10  $\mu$ M with CC<sub>50</sub> of 46.6, 27.8 and 28  $\mu$ M, resulting in a selectivity index (SI) of 4.88, 3.37 and 2.80, in addition to inhibition of 67.2, 69.7 and 69.3%, respectively. However, none of these compounds matched the selectioncriteria of a selective inhibitor of virus replication in this assay (i.e. >70% inhibition at concentrations that do not elicit an anti-metabolic effect on the host cells).

## 3.2.3 In vitro anti-Kinesin Eg5 activity

Antimitotic agents that have been used so far in cancer treatment, such as taxanes and vinca alkaloids, perturb tubulin polymerization/depolymerization, cause mitotic arrest and subsequent cell death [43]. However, these drugs produce serious side effects because microtubules also have essential intracellular functions in non-dividing cells [44]. A new alternative approach to prevent mitotic-spindle formation is the inhibition of proteins, such as the mitotic motors (kinesins) that interact with microtubules and cause mitotic arrest [45].

Monastrol (MA) has been reported as one of these antimitotic agents arrests cells in mitosis by specifically inhibiting Eg5 [46], a member of the Kinesin-5 family. Like many enzyme inhibitors, monastrol might be substrate competitive, inhibiting the ATP hydrolysis cycle of Eg5 by directly competing with ATP [45,47,48], or microtubule binding. Alternatively, monastrol might inhibit the motor domain allosterically, either by inhibiting ATP hydrolysis or by uncoupling partner head interactions to inhibit motor but not ATPase activity [49,50].

Compounds 17-28 have been selected for their inhibition of Eg5 activity using an *In vitro* malachite green ATPase assay (enzyme-coupled assay) [51]. None of these compounds matched the selection criteria of a selective inhibitor of Eg5 in this assay in comparison to monastrol, except 27 which showed ATPase inhibition value of 48% at 100  $\mu$ Mconcentration and inhibition of kinesin Eg5 with IC<sub>50</sub> of 4.87  $\mu$ M. From the data, it appears that a trifluoromethyl group on the 4-position of aromatic ring is important for Eg5 inhibition.

## 4. CONCLUSION

In this paper we have reported the synthesis and anti-HIV-1 and anti-HIV-2 evaluation of some indomethacin derivatives having bi- and triaryl groups as well as thioureido and amido residues of some l-amino acids. The preliminary *In vitro* anti-HIV activity of these analogues are too limited to perform extensive mode-of-action studies, however, 31 and 36 might be a new lead in the development of antiviral agents as nonnucleoside reverse transcpriptase inhibitors (NNRTIs). In addition, we reported the *In vitro* anti-HCV activity of three analogues but did not showed a selection inhibition of virus replication. Our study included the inhibition study of kinesin Eg5 *via* the ATPase activity assay, and the result indicated that 27, the only analouge of the indomethacin series showed moderate inhibition of ATPase activity (48%) at concentration of 100  $\mu$ M.

## **COMPETING INTERESTS**

Authors have declared that no competing interests exist.

## REFERENCES

- Hart F, Boardman P. Indomethacin: A New Non-steroid Anti-inflammatory Agent. Br Med J. 1963;2:965-970.
- 2. Shen TY, Winter Ch A. Chemical and biological studies of indomethacin. sulindac and their analogs. Adv Drug Res. 1977;12:90-245 (review).

- **3.** Humber LG, Ferdinandi E, Demerson CA, Ahmed S, Shah U, Mobilio D, Sabatucci J, De Lange B, Labbadia F, Hughes P. J Med Chem. 1988;31:1712-1719.
- 4. Moore PF, Larson DL, Otterness IG, Weissman A, Kadin SB, Sweeney FJ, et al. Tenidap, a structurally novel drugfor the treatment of arthritis: anti-inflammatory and analgesic properties. Inflamm Res. 1996;45:54-61.
- 5. Clària J. Cyclooxygenase-2 Biology. Curr Pharma Design. 2003;9:2177-2190 (review).
- Smith CJ, Zhang Y, Koboldt CM, Muhammad J, Zweifel BS, Shaffer A, et al. Pharmacological analysis of cyclooxygenase-1 in inflammation Proc Natl Acad Sci USA. 1998;95:3313-3318.
- 7. Ferreira S, Moncada S, Vane JR. Indomethacin and aspirin abolish prostaglandin release from the spleen. Nat New Biol. 1971;231:237-239.
- Wallace JL, McKnight W, Miyasaka M, Tamatani T, Paulson J, Anderson DC, et al. Role of endothelial adhesion molecules in NSAID-induced gastric mucosal injury. Am J Physiol. 1993;265:993-998.
- 9. Blobaum AL, Marnett LJ. Structural and functional basis of cyclooxygenase inhibition. J Med Chem. 2007;50:1425-441.
- 10. Prusakiewicz JJ, Felts AS, Mackenzie BS, Marnett LJ. Molecular Basis of the TimeDependent Inhibition of Cyclooxygenases by Indomethacin. Biochemistry. 2004;43:15439-15445.
- Felts AS, Ji C, Stafford JB, Crews BC, Kingsley PJ, Rouzer A, et al. Desmethyl Derivatives of Indomethacin and Sulindac as Probes for Cyclooxygenase-Dependent Biology. ACS Chem Biol. 2007;2:479-483.
- 12. Blobaum AL, Felts AS, Crews BC, Rouzer CA, Marnett L.J. The 2'-Trifluoromethyl Analogue of Indomethacin is a Potent and Selective COX-2 Inhibitor. ACS Med ChemLett. 2013;4:486-490.
- 13. Chennamaneni S, Zhong B, Lama R, Su B. COX inhibitors Indomethacin and Sulindac derivatives as antiproliferative agents: synthesis, biological evaluation, and mechanism investigation. Eur J Med Chem. 2012;56:17-29.
- 14. Meade EA, Smith WL, DeWitt DL. Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal antiinflammatory drugs. J Biol Chem. 1993;268:6610-6614.
- Boyer D, Bauman JN; Walker DP, Kapinos B, Karki K, Kalgutkar AS. Utility of MetaSite in improving metabolic stability of the neutral indomethacin amide derivative and selective cyclooxygenase-2 inhibitor 2-(1-(4-chlorobenzoyl)-5-methoxy-2-methyl-1H-indol-3-yl)-N-phenethyl-acetamide. Drugs Metab Dispos. 2009;37:999-; ref. therein in cited.
- Kalgutkar AK, Crews BC, Saleh S, Prudhomme D, Marnett L. Indolyl esters and amides related to indomethacin are selective COX-2 inhibitors. J Bioorg Med Chem. 2005;13:6810-6822.
- Chowdhury MA, Haung Z, Abdellatif KR, Dong Y, Yu G, Velazuez CA, Knaus EE. Synthesis and biological evaluation of indomethacin analogs possessing a Ndifluoromethyl-1,2-dihydropyrid-2-one ring system: a search for novel cyclooxygenase and lipoxygenase inhibitors Bioorg Med Chem Lett. 2010;20:5776-5780.
- Rajic Z, Butula I, Zorc B, Pavelic SK, Hock K, Pavelic K, Naesens L, De Clercq E, et al. Cytostatic and antiviral activity evaluations of hydroxamic derivatives of some nonsteroidal anti-inflammatory drugs. Chem Biol Drug Des. 2009;73:328-338.
- Camaco-Camaco C, Rojas-Oviedo L, Paz-Sandoval MA, Cardenas J, Gielen AT, Sosa LB, et al. Synthesis, structural characterization and cytotoxic activity of organotin derivatives of indomethacin App Organomet Chem. 2008;22:171-176.

- 20. Thun MJ, Henley SJ, Patrono C. Nonsteroidal anti-inflammatory drugs as anticancer agents: mechanistic, pharmacologic and clinical issues Natl Cancer Inst. 2002;94:252-266.
- 21. Piazza GA, Keeton AB, Tinsley HN, Whitt JD, Gary BD, Mathew B, et al. NSAIDs: Old Drugs Reveal New Anticancer Targets. Pharmaceuticals. 2010;3:1652-1667 (review).
- 22. Pavelici SK, Sedic M, PoznicM,Rajic Z, Zorc B, Pavelic K, Balzarin J, Mintas M. Evaluation of *In Vitro* Biological Activity of O-Alkylated Hydroxamic Derivatives of Some Nonsteroidal Anti-inflammatory Drugs Anticancer Res. 2010;30:3987-3994.
- 23. Amici C, Di CoroA, Ciucci A, Chiappa L, Castilletti C, Martella V, et al. Indomethacin has a potent antiviral activity against SARS coronavirus. AntivirTherap. 2006;11:1021-1030.
- 24. Richardson JY, Ottolini MG, Pletneva L, Boukhvalova M, Zhang S, Vogel SN, et al. Respiratory syncytial virus (RSV) infection induces cyclooxygenase 2: a potential target for RSV therapy. J Immunol. 2005;174:4356-4364.
- 25. Mukherjee PK, Simpson RW. Indomethacin inhibits viral RNA and protein synthesis in cells infected with vesicular stomatitis virus. Virology. 1984;140:188-191.
- 26. BourinbaiarAS, Lee-Huang S. The activity of plant-derived antiretroviral proteins MAP30 and GAP31 against herpes simplex virus in vitro. BiochemBiophys Res Commun. 1995;208:779-785.
- Al-Masoudi NA, Jafar NNA, Abbas LJ, Baqir SJ, Pannecouque C. Synthesis and anti-HIV Activity of New Benzimidazole, Benzothiazole and Carbohyrazide Derivatives of the anti-Inflammatory Drug Indomethacin.Z Naturforsch. 2011;66b:953-960.
- Pauwels R, Balzarini J, Baba M, Snoeck R, Schols D, Herdewijn P, Desmyter J, De Clercq E. Rapid and automated tetrazolium-based colorimetric assay for detection of anti-HIV compounds, J. Virol Methods. 1988; 20:309-321.
- 29. Pannecouque C, Daelemans D, De Clercq E. Tetrazolium-based colorimetric assay for the detection of HIV replication inhibitors: revisited 20 years later Nat. Protoc. 2008;3:427-434.
- Popovic M, Sarngadharan MG, Read E, Gallo RC. Detection, isolation, and continuous production of cytopathic retroviruses (HTLV-III) from patients with AIDS and pre-AIDS Science. 1984;224:497-500.
- 31. Barré-Sinoussi F, Chermann JC, Rey F, et al. Isolation of a T-lymphotropic retrovirus from a patient at risk for acquired immune deficiency syndrome (AIDS)Science. 1983;220:868-871.
- 32. Miyoshi I, Taguchi H, Kobonishi I., et al. Type C Virus-producing cell lines derived from adult T cell leukemia. Gann Monogr Cancer Res. 1982;28:219-228.
- 33. Vrolijk JM, Kaul A,Hansen BE, Lohmann V, Haagmans BL, Schalm SW BartenschlagerRA. A replicon-based bioassay for the measurement of interferons in patients with chronic hepatitis C. J Virol Methods. 2003;110:201-209.
- 34. Suzuki A. Syntheticstudies via the cross-couplingreaction of organoboronderivatives withorganichalides Pure AppChem. 1991;63:419- 422 (review).
- 35. Selvakumar K, Zapf A, Beller M. New Palladium-Carbene Catalysts for the Heck Reaction of Aryl Chlorides in Ionic Liquids. Org Lett. 2002;4:3031-3033.
- Kabbani AT, Ramadan H, Hammud HH, Ghannoum AM, Mouneimne YJ. Synthesis of some metal complexes of *N*-(benzoylamino)-thioxomethyl]amino acid (HL). UniChemTechn Metal. 2005;40:339-344.
- 37. Al-Masoudi NA, Al-Soud YA, Geyer A. <sup>1</sup>H- and <sup>13</sup>C-NMR study of some 6,7dihaloquinolone nucleosides and their derivatives. Spectroscopy Lett.1998;31:1031-1038.
- 38. Willker W, Leibfritz D, Kerssebaum R, Bermel W. Gradient selection in inverse heteronuclear correlation spectroscopy Mag Reson Chem. 1993;31:287-292.

- 39. Davis AL, Keeler J, Laue ED, Moskau D.Expwriments for recording pureabsorptionheteronuclear correlation spectra using pulsed field gradients J MagnReson. 1992;98:207-216.
- 40. Hargrave KD, Proudfoot JR, Grozinger KG, Cullen E, Kapadia SR, Patel UR, et al. Novel non-nucleoside inhibitors of HIV-1 reverse transcriptase. 1. Tricyclic pyridobenzo- and dipyridodiazepinones. J Med Chem. 1991;34:2231-2241.
- 41. Mitsuya H, Weinhold KJ, Furman PA, Clair MHSt, Lehrmann SN, Gallo, R. 3'-Azido-3'deoxythymidine (BW A509U): an antiviral agent that inhibits the infectivity and cytopathic effect of human T-lymphotropic virus type III/lymphadenopathy-associated virus *In vitro*. Proc Natl Acad Sci USA. 1985;82:7096-7100.
- 42. Janssen PAJ, Lewi PJ, Arnold E, Daeyaert F, de Jonge M, Heeres J. In search of a novel anti-HIV drug: multidisciplinary coordination in the discovery of 4-[[4-[[4-[[4-[(1E)-2-cyanoetenyl]-2-6-dimethylphenyl]amino]-2-pyrimidinyl]amino]-benzonitrile (R278474, Rilpivirine). J Med Chem. 2005;48:1901-1909.
- 43. Bhalla KN. Microtubule-targeted anticancer agents and apoptosis.Oncogene. 2003;22:9075-9086.
- 44. Jaffrezou JP, Dumontet WB, Derry G, Duran G, Chen E, Tsuchiya, et al. Novel mechanism of resistance to paclitaxel (Taxol) in human K562 leukemia cells by combined selection with PSC 833. Oncol Res. 1995;7:517-527.
- 45. Sharp DJ, Rogers GC, Scholey JM. Microtubule motors in mitosis. Natur. 2000;407:41-47.
- 46. Maliga Z, Kapoor TM, Mitchison T. Evidence that monastrol is an allosteric inhibitor of the mitotic kinesin Eg5. J Chem Biol. 2002;9:989-996.
- 47. Mayer TU, Kapoor TM, Haggarty SJ, King RW, Schreiber SL, MitchisonTJ.Small molecule inhibitor of mittotic spindle bipolarity identified in a phenotype-based screen. Science. 1999;286:971-974.
- Cochran JC. Gilbert SP. TPase Mechanism of Eg5 in the Absence of Microtubules: Insight into Microtubule Activation and Allosteric Inhibition by Monastrol. Biochemistry. 2005;44:16633-1648.
- 49. Cochran JC,Krzysiak TC, Gilbert SP. Pathway of ATP Hydrolysis by Monomeric Kinesin Eg5. Biochemistry. 2006;45:12334-12344.
- 50. Goulet A, Behnke-Parks WM, Sindelar CV, Major J, Rosenfeld SS, Moores C. J Biol Chem. 2012;287:44654-44666.
- 51. Henkel RD, VandeBerg JL, Walsh RA. A microassay for ATPase. Anal. Biochem. 1988;169:312-318.

© 2014 Al-Masoudi and Ali; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/3.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: http://www.sciencedomain.org/review-history.php?iid=424&id=16&aid=3870