



## **Inhibition of Microsomal Lipid Peroxidation and Protein Oxidation by *Carica papaya* (L) Leaf against Carbon Tetrachloride-Induced Hepatic Injury in Wistar Albino Rats**

O. J. Sule<sup>1\*</sup>, K. Kiridi<sup>2</sup> and A. R. Abdu<sup>3</sup>

<sup>1</sup>Department of Biochemistry, Faculty of Basic Medical Sciences, College of Health Sciences, Niger Delta University, Wilberforce Island, Bayelsa State, Nigeria.

<sup>2</sup>Department of Radiology, Faculty of Clinical Sciences, College of Health Sciences, Niger Delta University, Wilberforce Island, Bayelsa State, Nigeria.

<sup>3</sup>Department of Medical Microbiology and Parasitology, Faculty of Basic Medical Sciences, College of Health Sciences, Niger Delta University, Wilberforce Island, Bayelsa State, Nigeria.

### **Authors' contributions**

This work was carried out in collaboration between all authors. Author OJS designed and wrote the first manuscript of the study. Author KK wrote and performed the histopathological protocol. Author ARA performed the statistical analysis, analyses the study and conduct the literature searches. All authors read and approved the final manuscript.

### **Article Information**

DOI: 10.9734/EJMP/2016/13730

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Complete Peer review History: <http://sciencedomain.org/review-history/14748>

**Original Research Article**

**Received 1<sup>st</sup> September 2014**

**Accepted 28<sup>th</sup> April 2016**

**Published 23<sup>rd</sup> May 2016**

### **ABSTRACT**

**Aims:** To investigate the hepatoprotective activities of *C. papaya* leaf in Carbon tetrachloride induced hepatic damage in albino rats.

**Place and Duration of Study:** Department of Medical Biochemistry, Faculty of Basic Medical Sciences, College of Health Sciences, Niger Delta University, Wilberforce Island, Amassoma, Bayelsa-State, Nigeria, between April 2009 through July 2009.

**Methods:** Activity of total protein and bilirubin (direct and indirect bilirubin) in the serum were

\*Corresponding author: E-mail: J\_Sule@yahoo.com;

determined colourimetrically, while lipid peroxidation products, malondialdehyde (MDA), were measured in liver homogenate. Histopathological studies of the liver in both pretreated rats and controls were also, carried out. Group 1 (negative control) were fed with 100% rat feed. Groups 2-4 were pretreated with 10, 30 and 50% *C. papaya* leaf respectively, while Group 5 (normal control) received 100% rat feed. Rats in Groups (1-4), were injected with CCl<sub>4</sub> (0.5 ml/kg body weight in 0.5 ml olive oil) on the 29<sup>th</sup> day while rats in group 5 were not administered with CCl<sub>4</sub> (normal control). The results were statistically analyzed using one-way analysis of variance (ANOVA).

**Results:** There was significant increase ( $p < 0.05$ ) in the levels of malondialdehyde (MDA) and bilirubin (direct and indirect) in rats Group 1 when compared with the normal control. However, there is a significant decrease in the levels of these biochemical parameters (malondialdehyde and bilirubin) in the pretreated Groups (2-4) when compared with the negative control. Also, the protein levels significantly decreased ( $p < 0.05$ ) in the untreated rat Group 1 when compared the normal control. Pretreatment with *C. papaya* leaf significantly increased the levels of total protein in the rat Groups (2-4) when compared with the untreated Group 1. The decrease in the levels of MDA and bilirubin in the pretreated groups were concentration dependent. Rats in Group 4 that pretreated with 50% *C. papaya* leaf had the lowest values for MDA and bilirubin. Histopathology of the liver showed reduced level of injury with normal architecture in pretreated rats while; those not pretreated were presented with severe degrees of injuries.

**Conclusion:** Conclusively, the study reveals that *Carica papaya* may confer hepatoprotectivity to the rats exposed to carbon tetrachloride-induced liver damage.

**Keywords:** Histopathological; carbon tetrachloride; hepatoprotection; *C. papaya* leaf.

## 1. INTRODUCTION

Papaya is a man's common fruit and has a high nutritive value. Papaya is a polygamous species and it is difficult to identify a plant whether it is a male, female or hermaphrodite. The papaya tree belongs to a small family -*Caricaceae*, having four genera in the world. The genus *Carica* Linn. is represented by four species, of which *Carica papaya* Linn is the most widely cultivated and best known species. Papaya probably originated from Southern Mexico and Costa Rica, which was subsequently introduced as a plantation crop in Australia, Hawaii, Philippines, Sri Lanka, South Africa, India and in all tropical and subtropical regions [1]. It is grown for both commercial purpose and in home gardens. It is a tree reaching 3-10 m in height, with the habit of a palm; the fleshy stem marked by scars where leaves have fallen off, is surmounted by a terminal panache of leaves on long petioles and with 5-7 lobes [1]. The fruit bearing trees are less than 18 month old. The leaves and unripe fruits have been reported to contain milky juice in which the protein ferment Papain is presents [2,3]. It is low in calories and rich in natural vitamins and minerals. Papaya ranks among the first fruits for vitamin C, vitamin A, riboflavin, foliate, calcium, thiamine, iron, niacin, potassium and fibre [1]. Papaya has been reported also to contain broad spectrum of polychemicals including, polysaccharides, vitamins, minerals, enzymes, proteins, alkaloids, glycosides, fats

and oils, lectins, saponins, flavonoids, and steroids [4-8]. It was reported to exhibit therapeutic properties against various pathological disorder including tumours [9] and immunodeficiency which was based on the free radical scavenging activity as well as normalisation of an organism's super oxide level [10]. The fruit is rich in vitamins, minerals, proteins, polysaccharides, lectins, saponins and flavonoids, and can be used in the prevention of complications of diabetes mellitus [11]. The black seeds are edible and have a sharp, spicy taste. They are sometimes ground up and used as a substitute for black pepper. In some parts of Nigeria, the young leaves of papaya are steamed and eaten like spinach. Hence, the reason for this study is to investigate the biochemical effects of *C. papaya* leaf on the carbon tetrachloride-induced liver of wistar albino rats.

## 2. MATERIALS AND METHODS

### 2.1 Preparation of Papaya Leaf Powder

Fresh samples of *C. papaya* leaves were collected from the herbal garden of Department of Pharmacognosy, Faculty of Pharmacy, Niger Delta University, Wilberforce Island, Bayelsa State, Nigeria. The plant was identified and authenticated by a Taxonomist, Prof S.K. Adesina of Department of Pharmacognosy and was deposited in the herbarium with voucher no: NDUP 093. The leaves were detached from the

stems, washed twice with distilled water to remove adulterants, dried under natural conditions for two weeks, ground into powder using an electric blender and stored in airtight containers and used within duration of study.

## 2.2 Animals

Thirty (30) male albino rats weighing (180-190 g) were obtained from Department of Pharmacology Animal House, Niger Delta University. The animals were housed in stainless-steel cages under standard laboratory conditions of  $27\pm 2^{\circ}\text{C}$ , relative humidity  $50\pm 15\%$  and normal photo period (12 h dark/12 h light). They were maintained on standard rat chow and water *ad libitum* and acclimatised for a period of seven days. Permission and approval for animal studies were obtained from College of Health Sciences, Animal Ethics Committee, Niger Delta University with Ref: No. NDU/CHS/FBMS/0078.

## 2.3 Phytochemical Screening

In 2008, Krishna et al. [1] reported the chemical composition of *C. papaya* to include; alkaloid,  $\alpha$ -carpaine,  $\beta$ -D-glucosides,  $\beta$ -sitosterol, papain, choline, carotene, riboflavin, vitamin C, phenylethyl-  $\beta$ - D- glucosides, amongst others.

## 2.4 Experimental Design

Thirty (30) albino rats (180-190 g) were randomly divided into five groups comprising of six rats each. Animals were fed with substances under investigation for a period of twenty eight (28) days after which they were injected with 0.5 ml/kg  $\text{CCl}_4$ , dissolved in 0.5 ml olive oil. They were fed *ad libitum* with free access to water. Group 1 (negative control) were fed with 100% rat feed. Groups 2- 4 were pretreated with 10, 30 and 50% *C. papaya* leaf respectively, while Group 5 (normal control) received 100% rat feed. Rats in Groups (1-4), were injected with  $\text{CCl}_4$  (0.5 ml/kg body weight in 0.5 ml olive oil) on the 29<sup>th</sup> day while rats in Group 5 were not administered with  $\text{CCl}_4$  (normal control).

## 2.5 Sample Collection

Twenty four hours after injection of  $\text{CCl}_4$ , the rats were anaesthetized in chloroform saturated chamber, sacrificed and blood samples were obtained through cardiac puncture, in non-heparinised tubes, centrifuged at 3000 rpm for 10 minutes and blood sera were then collected and stored at  $4^{\circ}\text{C}$  prior to immediate estimation of

total protein, bilirubin (direct and indirect bilirubin). The liver from both control and test animals were removed and weighed to the nearest 0.01 g. The livers were removed immediately, washed with ice-cold saline and a 10% homogenate was prepared in phosphate buffer (pH 7.0). The homogenate was centrifuged at 3000 rpm for 10 min at  $4^{\circ}\text{C}$  and the supernatant was used for the estimation of MDA. The pieces of liver were preserved in 10% formaldehyde solution for histological study.

## 2.6 Biochemical Assay

Determination of total protein was by colorimetric method (Biuret method), as modified by Giodani et al. [12]. Bilirubin was estimated by colorimetric method of Jendrassik and Grof [13] while, Lipid peroxidation products MDA was according to Hunter et al. [14] as modified by Gutteridge and Wilkins [15].

## 2.7 Histopathological Examination

The pieces of liver were preserved in 10% formaldehyde solution for histopathological examination according to the method of Baker and Silverton [16].

## 2.8 Statistical Analysis

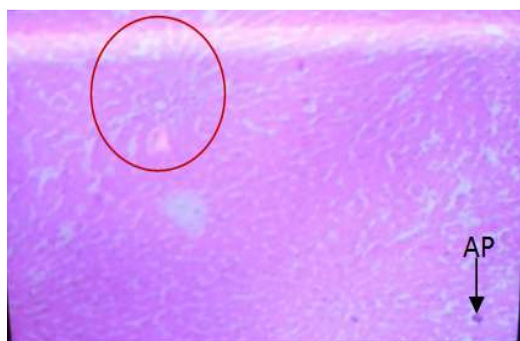
The results were statistically analysed using one-way analysis of variance (ANOVA) using SPSS version 17, followed by Tukey-Kramer Multiple Comparisons Test. P values  $< 0.05$  were considered significant.

## 3. RESULTS

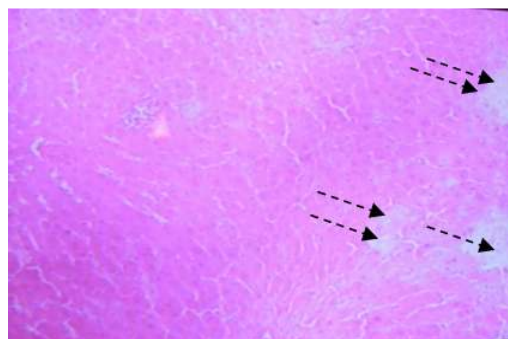
### 3.1 Effect of *C. papaya* Leaf on Some Biochemical Parameters

The results of some biochemical indices shown in Table 1, indicated a significant decrease ( $p < 0.05$ ) in the levels of total proteins in rats that were administered  $\text{CCl}_4$  only (Group 1), when compared with rats that were pre-treated with 10, 30 and 50% *Carica papaya* (Groups 2, 3 and 4 respectively). The levels of total proteins were significantly increased ( $p < 0.05$ ) in a dose dependent manner in rats that were pre-treated with 10, 30 and 50% *Carica papaya*. Rats administered with  $\text{CCl}_4$  only (Group 1), showed significant increase ( $p < 0.05$ ) in the levels of total bilirubin. However, pre-treatment with 10, 30 and 50% *Carica papaya* (Groups 2, 3 and 4 respectively) significantly decreased ( $p < 0.05$ ) total bilirubin levels in a dose dependent manner.

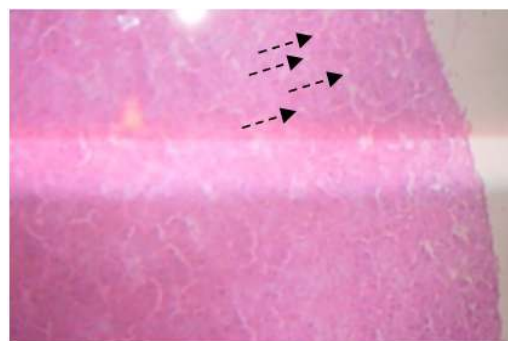
The levels of direct and indirect bilirubin significantly increased ( $p < 0.05$ ) in rat that were administered  $CCl_4$  only, when compared with the rats in Group 5 (normal control). Rats that were pre-treated with 10, 30 and 50% *Carica papaya* showed significant decrease ( $p < 0.05$ ) in the levels of direct and indirect bilirubin, when compared with rats that were administered with  $CCl_4$  only. Malondialdehyde (MDA) levels increased significantly ( $p < 0.05$ ) in rats injected with  $CCl_4$  only, when compared to rats that were not administered with  $CCl_4$  (Groups 5)). However there was significant decrease ( $p < 0.05$ ) in the levels of MDA in rats that were pre-treated with 10, 30 and 50% *Carica papaya* (Groups 2, 3 and 4 respectively), when compared with rats that were treated with  $CCl_4$  only.



**Plate 1.** Liver slide of rats administered 100% Feed +  $CCl_4$ : (negative control) Portal triaditis (mild) (red circle) and apoptosis (AP) of individual liver cells. H&E 100X



**Plate 2.** Liver slide of rats administered 10% *Carica papaya* +  $CCl_4$ : Mild microvesicular steatosis (dash arrows) of the liver. H&E 100X



**Plate 3.** Liver slide of rats administered 30% *Carica papaya* +  $CCl_4$ : Normal liver architecture with mild microvesicular steatosis (dash arrows). H&E 100X

**Table 1.** The effects of *C. papaya* leaf on some biochemical parameters in  $CCl_4$  induced hepatotoxicity

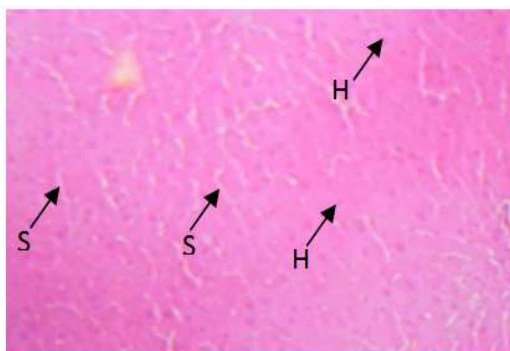
Group	Treatments	Total bilirubin ( $\mu\text{mol/L}$ )	Direct bilirubin ( $\mu\text{mol/L}$ )	Indirect bilirubin ( $\mu\text{mol/L}$ )	Total protein (g/L)	MDA $\mu\text{mol/L} \times 10^{-5}$
1	100% FEED + $CCl_4$	26.13 <sup>a</sup> ±0.20	14.76 <sup>a</sup> ±0.20	11.37 <sup>a</sup> ±0.20	40.25 <sup>a</sup> ±0.20	4.50 <sup>a</sup> ±0.01
2	90% FEED + 10% <i>C. papaya</i> leaf+ $CCl_4$	13.52 <sup>b</sup> ±0.10	7.38 <sup>b</sup> ±0.10	6.14 <sup>b</sup> ±0.10	52.96 <sup>b</sup> ±0.10	3.50 <sup>b</sup> ±0.01
3	70% FEED +30% <i>C. papaya</i> leaf+ $CCl_4$	10.50 <sup>c</sup> ±0.20	5.88 <sup>c</sup> ±0.20	4.62 <sup>c</sup> ±0.20	56.86 <sup>c</sup> ±0.10	3.20 <sup>c</sup> ±0.01
4	50% FEED + 50% <i>C. papaya</i> + $CCl_4$	9.90 <sup>d</sup> ±0.10	5.30 <sup>d</sup> ±0.10	4.60 <sup>c</sup> ±0.10	63.48 <sup>d</sup> ±0.10	2.70 <sup>d</sup> ±0.01
5	100% FEED - $CCl_4$ (General Control)	9.20 <sup>d</sup> ±0.10	5.20 <sup>d</sup> ±0.10	4.20 <sup>c</sup> ±0.10	72.76 <sup>e</sup> ±0.10	2.60 <sup>e</sup> ±0.01

Values are mean±S. D for 6 replicates (n= 6)

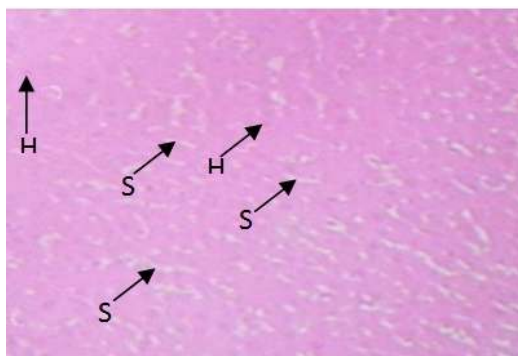
Means with different superscripts are significantly different at the 0.05 levels in the columns

### 3.2 Histopathological Examination

To ascertain the effects of various treatments on the organs of the rats, the histopathological examinations of liver of rats were carried out. Tissue slides of liver of rats in test and control groups were prepared and the results are as shown.



**Plate 4. Liver slide of rats administered 50% *Carica papaya* + CCl<sub>4</sub>: Normal liver architecture. Hepatocyte (H), Sinusoid (S) H&E 100X**



**Plate 5. Liver slide of rats administered 30% *Carica papaya* – CCl<sub>4</sub> (normal control): Normal liver architecture. Hepatocyte (H), Sinusoid (S) H&E 100X**

### 4. DISCUSSION

Hepatoprotective activity of *Carica papaya* was investigated using biochemical indices such as total protein, direct and indirect bilirubin and lipid peroxidation products. The results of this study suggests that rat groups that were pretreated with *Carica papaya* prior to administration of CCl<sub>4</sub> showed a significant decrease in hepatotoxicity when compared with rats in Group 1 (negative control). The results showed a significant

decrease in the levels of total bilirubin (direct and indirect) and malondialdehyde (MDA), in the serum. The estimation of these biochemical indices in the serum was reported to be a useful quantitative marker for the extent and type of hepatocellular damage [17]. The results of this study further agree with the earlier observation of Babalola et al. [18], that the terpenoid fraction of *V. amygdalina* leaf extract ameliorates carbon tetrachloride induced hepatotoxicity in rats. Krishna et al. [1] earlier reported the chemical composition of *Carica papaya* to include; alkaloid,  $\alpha$ -carpaine,  $\beta$ -D-glucosides,  $\beta$ -sitosterol, papain, choline, carotene, riboflavin, vitamin C, phenethyl-  $\beta$ - D- glucosides, amongst others. These compounds have been reported to have the potentials to function as antioxidants by scavenging the superoxide anion, hydroxyl radical and peroxy radical or quenching singlet oxygen thus inhibiting lipid peroxidation in biological system [19]. *C. papaya* leaf was also reported to exhibit potential supportive role on oxidative inflammatory damage in cirrhosis caused by hepatitis C virus [20]. It was also suggested by Sallie et al. [21] that the reduction of serum bilirubin levels in pretreated rat groups (Groups 2, 3 and 4 in Tables 1), may provide scientific justification for its use by herbalists as a treatment for jaundice being most sensitive markers employed in the diagnosis of hepatic damage because they are released into circulation after cellular damage. The results also showed an increased serum total protein levels in rats pretreated with *Carica papaya* (Table 1), which could be an indication that the synthetic function of the liver was improved. A decrease in serum total proteins was earlier reported as indications of hepatotoxicity [22]. Histopathology of the liver cells revealed portal triaditis (mild) and apoptosis of individual cells in untreated rat Group 1, indicating some levels of cellular damage due to toxicity, hence supporting the biochemical results. Mild microvesicular steatosis of individual liver cells with normal architecture was also observed in rats pretreated with 10% *Carica papaya*. However, normal liver architecture with mild microvesicular steatosis especially beneath the capsular area was seen in rats pretreated with 30% *Carica papaya*. The livers of rats pretreated with 50% *Carica papaya* and normal control groups showed normal architecture and did not indicate any damages as reported in Plates 4 and 5 respectively. This is in agreement with the reports by Aravind et al. [23] that *C. papaya* leaf could reduce the risk of disease caused by free radical activities and high cholesterol in the blood [24]. Thus, the study

reveals that *Carica papaya* may confer hepatoprotectivity to the rats exposed to carbon tetrachloride-induced liver damage.

## 5. CONCLUSION

Conclusively, the study suggests that *Carica papaya* may possess hepatoprotectivity to the rats exposed to carbon tetrachloride-induced liver damage.

## CONSENT

It is not applicable.

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

## REFERENCES

1. Krishna KL, Paridhavi M, Jagruti AP. Review on nutritional, medicinal and pharmacological properties of papaya (*Carica papaya* Linn.). Nat. Prod. Radiance. 2008;7(4):364-373.
2. Bungornm S, Varima W, Pisamai L, Jamsai S, Dusit J. Diuretic effects of selected Thai indigenous medicinal plants in rats. J Ethnopharmacol. 2001;75(2-3): 185-190.
3. Bhattacharjee SK. *Carica papaya*. In Shashi J, editor. Handbook of medicinal plants. 3<sup>rd</sup> Revised ed. Jaipur: Pointer Publisher; 2001.
4. Roy SD, Chakraborty J, Shil D, Das S, Begum N. Herbs used in peptic ulcer: A Review. IJPRAS. 2013;2(2):9-23.
5. Nahak G, Suar M, Sahu RK. Antioxidant potential and nutritional values of vegetables: A review. Res. J. Med. Plant; 2013. DOI: 10.3923/rjmp.2013
6. Sule OJ, Elekwa I, Joffa PPK. Morphological and biochemical effects of dried leaves of *Carica papaya* Linn. (Pawpaw) on the liver in wistar rats. J Pharm Biomed Sci. 2012;15(15):1-5. Available: [www.jpbums.info](http://www.jpbums.info)
7. Kothari V, Seshadri S. Antioxidant activity of seed extracts of *Annona squamosa* and *Carica papaya*. J. Nutr Food Sci. 2010; 40(4):403-408.
8. Bhattacharjee SK. *Carica papaya* in: Hand book of medicinal plants, 3rd Revised Edn, by Shashi Jain (Ed), Pointer Publisher, Jaipur. 2001;1-71.
9. Otsuki N, dang DH, Kumaqai E, Kondo A, Iwata S, Morimoto C. Aqueous extract of *Carica papaya* leaves exhibit anti-inflammatory activity and immunomodulatory effects. J Ethnopharmacol. 2010;127(3):760-767.
10. Osanto JA, Santiago LA, Remo GM, Cuadra MS, Mori A. Antimicrobial and antioxidant activities of unripe papaya. Life Sci. 1993;53(17):1383-1389.
11. Savickiene N, Dagilyte A, Lukosius A, Zitkevicius V. Importance of biologically active components and plants in the prevention of complications of diabetes mellitus. Medicina (Kaunas). 2002;38(10): 970-975.
12. Giodani R, Cardenas ML, Moulin-Traffort J, Regli P. Fungicidal activity of latex sap from *Carica papaya* and antifungal effect of D (+) glucosamine on *Candida albicans* growth. Mycoses. 1996;39(3-4):103-110.
13. Jendrassik L, Grof P. Vereinfachte photometrische methode zur beshmung des bilirubins. Biochem Z. 1938;297:81-89.
14. Hunter MIS, Mohammed JB. Plasma antioxidants and lipid peroxidation products in duchenne muscular dystrophy. Clinton. Chin Acts. 1986;155:123-132.
15. Gutierrez JMC, Wilkins C. Copper dependent hydroxyl radical damage to ascorbic acid formation of a thiobarbituric acid reactive products. FEBs. LEH. 1982;137:327-340.
16. Baker JF, Silverton ER. Kishaw D. Introduction to medical laboratory technology. London: Butterworths; 1985.
17. Ansari RA, Tripathi SC, Patnaik GK, Dhawan BN. Antihepatotoxic properties of picroliv, an active fraction from rhizomes of *Picrorhiza kurroa*. J. Ethnopharmacol. 1991;34:61-68.
18. Babalola OO, Anetor JI, Adeniyi FA. Amelioration of carbon tetrachloride-induced hepatotoxicity by terpenoid extract from leaves of *Vernonia amygdalina*. Afr J Med Med Sci. 2001;30:91-93.
19. Severi JA, Lima ZP, Kushima H, Brito LC, dos Santos ARMS, Vilegas W, Hiruma-Lima CA. Polyphenols with antiulcerogenic action from aqueous decoction of mango leaves (*Mangifera indica* L.). Molecules. 2009;14:1098-1110. DOI: 10.3390/molecules 14031098

20. Marotta F, Weksler M, Naito Y, Yoshida C, Yoshioka M, Marandota P. Nutraceutical supplementation, effect of a fermented papaya preparation on redox status and DNA damage in healthy elderly individuals and relationship with GSTM1 genotype, a randomized, placebo-controlled, cross-over study. *Ann N Y Acad Sci.* 2006;1067(1):400-407.
21. Sallie R, Tredger RS, Williams R. Drugs and the liver. *Biopharm. Drug Dispos.* 1991;12:251-259.
22. Abatan MO, Arowolo ROA, Olorunsogo O. Pathological effects of *Lantana camara* and *Dichapetalum madagascasiense* in goats. *Trop. Vet. Med.* 1996;14:127-132.
23. Aravind G, Debjit B, Duraivel S, Harish G. Traditional and medicinal uses of *Carica papaya*. *J. Med. Plants Stud.* 2013;1(1):7-15.
24. Senthilkumaran JJ, Shalini N. An overview of *Carica papaya* and its medicinal uses. *Res J Pharm Biol Chem Sci.* 2014;5(2): 641-649.

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